

Review

Comparative Histomorphological Review of Rat and Human Hepatocellular Proliferative Lesions

Bob Thoolen^{1,2*}, Fiebo J.W. ten Kate², Paul J. van Diest², David E. Malarkey³, Susan A. Elmore³, and Robert R. Maronpot⁴

¹Global Pathology Support, Benoordenhoutseweg 23, 2596 BA The Hague, The Netherlands

²University Medical Center Utrecht, PO Box 85500, 3508 GA Utrecht, The Netherlands

³National Toxicology Program, National Institute of Environmental Health Sciences, Cellular and Molecular Pathology Branch, 111 T.W. Alexander Drive, NC 27709, USA

⁴Maronpot Consulting LLC, 1612 Medfield Road, Raleigh, NC 27607, USA

Abstract: In this comparative review, histomorphological features of common nonneoplastic and neoplastic hepatocyte lesions of rats and humans are examined using H&E-stained slides. The morphological similarities and differences of both neoplastic (hepatocellular carcinoma and hepatocellular adenoma) and presumptive preneoplastic lesions (large and small cell change in humans and foci of cellular alteration in rats) are presented and discussed. There are major similarities in the diagnostic features, growth patterns and behavior of both rat and human hepatocellular proliferative lesions and in the process of hepatocarcinogenesis. Further study of presumptive preneoplastic lesions in humans and rats should help to further define their role in progression to hepatocellular neoplasia in both species. (DOI: 10.1293/tox.25.189; *J Toxicol Pathol* 2012; 25: 189–199)

Key words: liver, hepatocellular carcinoma, hepatocellular adenoma, large cell change, small cell change, foci of cellular alteration

Introduction

Primary hepatocellular carcinoma (HCC) is the fifth most common cancer in the world and the third most frequent cancer-related cause of death with increasing incidence worldwide^{1–4}. In addition, HCC is the most common primary liver malignancy in the world^{5–7}. In the majority of cases, it is associated with hepatitis B or C viral infections, aflatoxicosis, and/or liver cirrhosis^{8–11}. Other risk factors for developing HCC include alcoholic liver disease, nonalcoholic steatohepatitis, diabetes, and obesity^{12,13}. Most patients with HCC are diagnosed at a late stage; therefore, the prognosis of HCC patients is generally very poor, with a 5-year survival rate of less than 5%^{7,14}.

Experimental rat and mouse hepatocarcinogenesis models have been used for decades to delineate the pathogenesis of hepatic neoplasia. The rodent experimental model is used to identify potential human carcinogenic risk from exposure to drugs, environmental agents, and other xenobi-

otics. Rat hepatocellular adenomas (HCAs) and carcinomas are commonly used in tumor response and carcinogenicity bioassays and share some common features with human adenomas and carcinomas¹⁵.

In rat experimental models, presumptive preneoplastic foci of cellular alteration occur prior to the appearance of hepatocellular adenomas and HCC; however, there is experimental evidence that not all foci of cellular alteration progress to neoplasia and that some may actually regress^{16,17}. Basophilic (BAS), eosinophilic (EOS), and clear cell (CLEAR) foci of cellular alteration in rats are the counterparts of human liver cell dysplasias classified as large cell change and small cell change. The detection of these presumptive preneoplastic lesions in humans may be indicative of progression towards HCC^{18–21} although further investigation is warranted. The purpose of this overview is to compare and contrast the morphological features of representative examples of commonly occurring human and rat hepatoproliferative lesions and to report the biology of these lesions.

Method

Paraffin blocks of adult human cases were selected from the archives of the Departments of Pathology, University Medical Center Utrecht (UMCU) and Erasmus Medical Center Rotterdam, The Netherlands. These surgical specimens were reviewed and considered unequivocal examples

Received: 1 May 2012, Accepted: 24 May 2012

*Corresponding author: B Thoolen (e-mail: bob.thoolen@gpstoxpath.com)

©2012 The Japanese Society of Toxicologic Pathology

This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives (by-nc-nd) License <<http://creativecommons.org/licenses/by-nc-nd/3.0/>>.

Table 1. H&E-stained Human (UMCU) / Rat (NTP) Liver Lesions

Lesion	Human			Rat			
	n	Gender %		n	Gender%		
FNH	12	8	92	–	–	–	
HCC	16	88	12	15	20	80	
HCA	15	–	100	10	40	60	
Dysplastic lesions	n	M	F	Foci of cellular alteration	n	M	F
LCC	19 ¹	100	–	FCA/EOS	11	91	9
SCC	17 ²	78	22	FCA/BAS	9	89	11
				FCA/CLEAR	9	100	–

FNH, Focal Nodular Hyperplasia; HCA, Hepatocellular Adenoma; LCC, Large Cell Change; SCC, Small Cell Change; FCA, Focus of Cellular Alteration; EOS, Eosinophilic; BAS, Basophilic; CLEAR, Clear cell.

¹ From 10 different patients; gender % is based on 10 people with at least one lesion. ² From 9 different patients; gender % is based on 9 people with at least one lesion.

of human focal nodular hyperplasias (FNHs), HCCs, HCAs, large cell change (LCC) and small cell change (SCC).

Paraffin blocks of rat cases obtained from the National Toxicology Program (NTP) archives were from studies of chemical-induced liver tumors and represent diagnoses peer reviewed by experienced rodent toxicologic pathologists. Rat cases include HCCs, HCAs and basophilic, eosinophilic and clear cell foci of cellular alteration (FCAs). FNH lesions were not identified as they are rare in rats. However, this lesion was included in humans since it is one of the most common human proliferative liver lesions.

Original slides from the human and rat cases were reviewed by a medical liver pathologist and two toxicologic pathologists and selected using published diagnostic criteria^{8,22–26} to confirm original diagnoses. Once confirmed, additional sections for this study were prepared, and all hematoxylin and eosin (H&E) staining was performed simultaneously at the UMCU after collection of all unstained paraffin slides on coated glass slides (e.g., Superfrost Plus) (see Table 1).

Results

Human cases

1. Focal nodular hyperplasia (FNH)

Liver samples were derived from surgical excision (hemihepatectomy, partial liver resection or biopsy) of female patients (11/12; 92%) and one male patient (1/12; 8%) at the UMCU. The corresponding resection specimens included in the study for comparison, were grossly nodular and ranged in diameter from 2 to 17 cm. Microscopically, they had classical diagnostic features of FNH consisting of nodules composed of plates of hyperplastic hepatocytes that were two-cell layers thick and subdivided by fibrous septa (Fig. 1). Thick-walled arteries were present in the stellate scars and septa, and there were bile ductules typically located between the scars and the liver parenchyma (Fig. 2). Near the fibrous septa there were occasionally small immature cells with oval to fusiform leptochromatic nuclei and scant cytoplasm that resembled rat oval cells. In addition, tran-

sitional cells displaying characteristics of both hepatocytes and bile duct cells were also present in some samples. These results suggest the presence of “undifferentiated progenitor cells” within FNH and further suggests that the ductular reaction, at least partly, can be explained by activation of these cells²⁷.

2. Hepatocellular carcinoma (HCC)

For the human samples, 14/16 HCC (88%) were from male patients. The morphological features consisted of a broad trabecular growth pattern of hepatocytes with occasional mixed growth patterns of trabecular/compact (Fig. 3), trabecular/acinar (Fig. 5) and sometimes a mixture of the three growth patterns. Hemorrhage, ischemic necrosis, neovascularization, angiectasis or peliosis hepatis and cystic changes were more commonly observed in these malignant tumors as compared to the other lesions evaluated.

3. Hepatocellular adenoma (HCA)

HCAs from female patients (n=15) had a maximum diameter of 16 cm. Histologically, they matched the common diagnostic criteria (see Table 2) for this benign liver neoplasm and sometimes showed focal to more diffuse steatosis, which can be observed in these tumors (Fig. 7)^{28–30}.

Human cases (continued)

1. Liver cell dysplasia – large cell change (LCC) and small cell change (SCC) (human)

Large cell change: LCC (synonyms: large liver cell change (LLCC) or large liver cell dysplasia (LLCD)) has been described in detail by Anthony *et al.*³¹ and others^{24,32–36}. Morphological features of hepatocytes with large cell change included cellular enlargement, nuclear pleomorphism with hyperchromasia, prominent nucleoli and occasional multinucleation. Enlargement was usually two- to three-fold and both nuclear and cytoplasmic with a normal nucleus to cytoplasmic ratio. Cytoplasmic staining was normal with occasionally more or less glycogen than that present in the surrounding liver parenchyma³¹. A classical example of such lesions is illustrated in Fig. 9. The selected cases of LCC (n=19) were all from 10 male patients.

Small cell change: SCC (synonyms: small liver

Table 2. Major Diagnostic Criteria for Proliferative Liver Lesions

Lesion	n	Definition
FNH	12	<ul style="list-style-type: none"> - Nodules of hyperplastic hepatocytes with two–cell layer hepatic plates divided by fibrous septa - Ductular reaction - Stellate scars with thick-walled arteries - Arising in a normal liver
HCC	16	<ul style="list-style-type: none"> - Trabecular or mixed growth patterns of atypical hepatocytes - Alteration of tinctorial staining patterns and marked cellular pleomorphism may occur - No portal tracts (unless entrapped by the normal liver) - Vascular/stromal invasion - Hemorrhage, necrosis, neovascularization, angiectasis and cystic changes more common than in HCA - Loss of normal reticulin framework - Isolated arteries - Mitotic index increased
HCA	15	<ul style="list-style-type: none"> - Proliferation of benign hepatocytes without acinar structures - Loss of normal lobular architecture - Compression of the surrounding parenchyma - Focal or diffuse steatosis - No more than 3 nodules/liver - Isolated arteries and arterioles - Mitotic index may be increased
Dysplastic lesions (human)		
LCC	19	<ul style="list-style-type: none"> - (Foci of) enlargement of hepatocytes 2- to 3-fold (both cytoplasmic and nuclear) - Nuclear pleomorphism with hyperchromasia - Prominent nucleoli - Multinucleation - Normal cytoplasm with less or more glycogen
SCC	17	<ul style="list-style-type: none"> - (Foci of) small hepatocytes with a high N:C-ratio - Nuclear atypia and different cytoplasmic staining - Fat or glycogen may differ from surrounding liver cells
Foci of cellular alteration (rat)		
FCA/EOS	11	<ul style="list-style-type: none"> - Normal or minimal compression of the surrounding parenchyma - (Foci of) enlarged, polygonal hepatocytes with (increased) acidophilic staining - Granular and pale intense eosinophilic cytoplasm of hepatocytes sometimes with a ground-glass appearance - Glycogen and/or some clear cells may be present
FCA/BAS	9	<ul style="list-style-type: none"> - Normal or minimal compression of the surrounding parenchyma - (Foci with) basophilic staining of hepatocytes of normal/smaller size - Cells sometimes arranged in tortuous cords and dissociation of cells may occur - Liver plates merge imperceptively with the surrounding parenchyma - Cells may be pleomorphic with enlarged (vesiculated) nuclei and prominent nucleoli
FCA/CLEAR	9	<ul style="list-style-type: none"> - Normal or minimal compression of the surrounding parenchyma - Normal/enlarged hepatocytes with acidophilic staining - Small nuclei, dense, centrally located - Excess storage of glycogen - Prominent cell membranes

FNH, focal nodular hyperplasia, HCA, hepatocellular adenoma, LCC, large cell change, SCC: small cell change, FCA: focus of cellular alteration; EOS, eosinophilic; BAS, basophilic; CLEAR, clear cell.

cell change (SLCC) or small liver cell dysplasia (SLCD)) was characterized by small hepatocytes with a high nuclear:cytoplasmic (N:C) ratio. Cells were uniform and differed from cells of the surrounding parenchyma in terms of nuclear atypia and cytoplasmic staining. Fat or glycogen content sometimes differed from that in the adjacent liver parenchyma. These collections of cells with small cell change tended to produce more small round distinct foci with irregular margins similar to foci that are more closely associated with the HCCs (Fig. 11) as reported by oth-

ers^{37–39}. The selected SCC cases (n=17) were from 9 patients (7/9, 78% males and 2/9, 22% females). In the cases evaluated, combined areas of LCC and SCC could sometimes be observed within the same slide.

Rat cases

1. Hepatocellular carcinoma (HCC)

The HCCs reviewed, were from dosed males (3/15 HCC, 20%) and females (12/15, 80%). The morphological features were consistent with published HCC criteria²⁶

and exhibited largely trabecular growth patterns, although mixed growth patterns (trabecular/acinar/solid) and occasionally basophilic and eosinophilic areas were present. In one case, the HCC arose within a hepatocellular adenoma and had an infiltrative growth pattern. A trabecular growth

pattern with focal steatosis and mitosis is illustrated in Figure 4. As was seen in human HCCs, sometimes areas with acinar growth patterns were observed (Fig. 6). Most lesions evaluated also had hemorrhage, necrosis, pigment deposition (hemosiderin), angiectasis and/or focal fatty change.

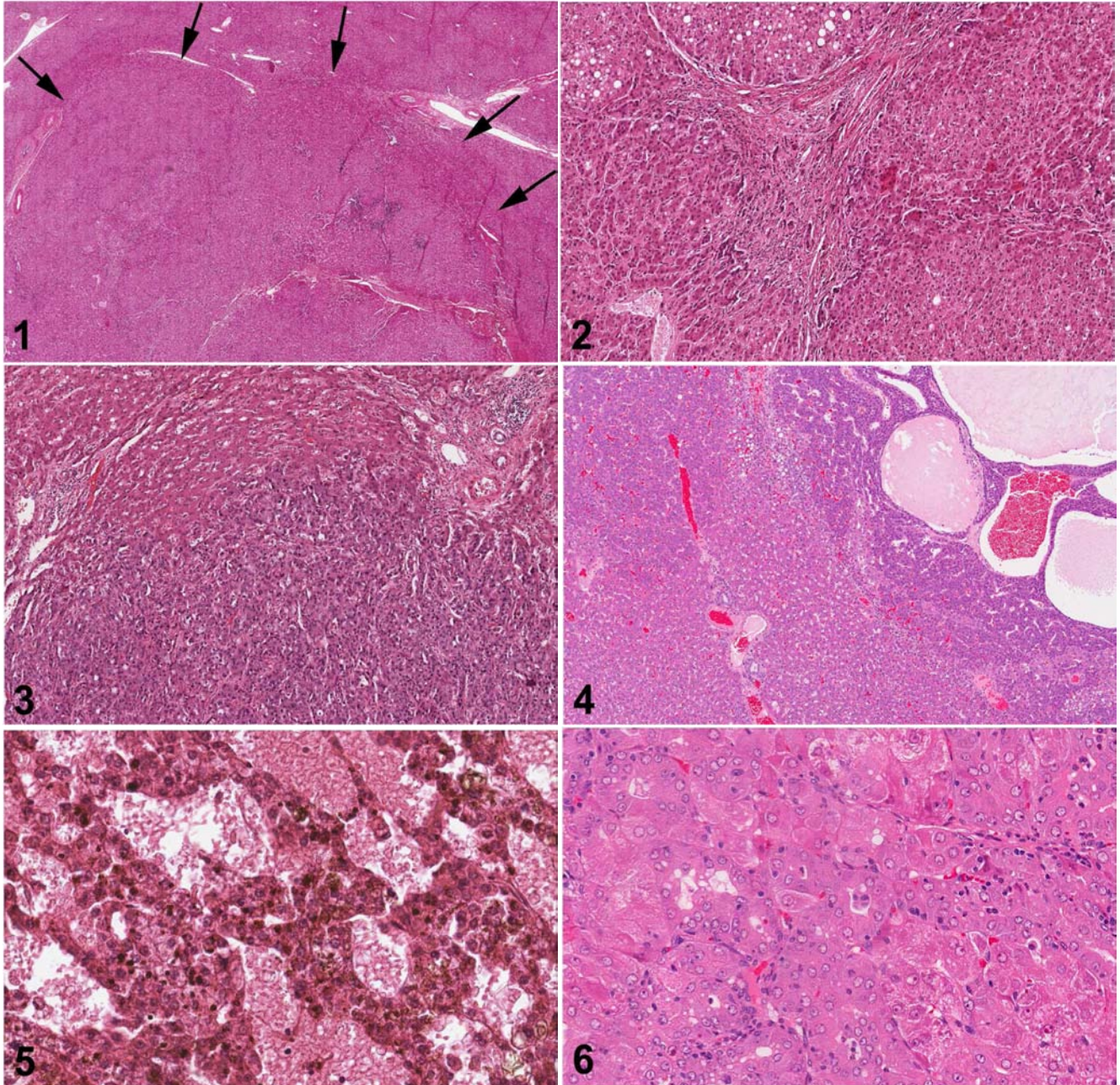


Fig. 1. Human liver. Low magnification of FNH. The upper border of the FNH is indicated by arrows with relatively normal hepatic parenchyma at the top of the figure. H&E.

Fig. 2. Human liver. Higher magnification of Fig. 1. The FNH consists of nodules composed of two-cell layers of hepatocytes subdivided by fibrous septa. Proliferative ductules are present in stellate septal scars. H&E.

Fig. 3. Human liver. Hepatocellular carcinoma composed of atypical hepatocytes arranged in a solid or trabecular growth pattern with normal hepatic parenchyma present at the top of the figure. H&E.

Fig. 4. Rat liver. Hepatocellular carcinoma with a trabecular growth pattern at the top and right of the figure. There is angiectasis in the carcinoma on the right. Normal hepatic parenchyma is present on the lower left of the figure. H&E.

Fig. 5. Human liver. High magnification of a hepatocellular carcinoma with a mixed acinar and trabecular growth pattern. H&E.

Fig. 6. Rat liver. High magnification of a hepatocellular carcinoma with a mixed acinar and trabecular growth pattern. H&E.

2. Hepatocellular adenoma (HCA)

The selected hepatocellular adenomas (n=10) were from animals of both sexes (6/10, 60% females and 4/10, 40% in males). The histological features were compatible with the common diagnostic criteria (see Table 2). Sometimes, these adenomas also contained diffuse fatty change (steatosis) and angiectasis (Fig. 8).

3. Focus of cellular alteration (FCA)

Eosinophilic foci: Eosinophilic foci of cellular alteration consisted of polygonal enlarged hepatocytes with increased acidophilic staining compared with the surrounding normal liver. Clear cells (glycogen storage) were occasionally present. The cytoplasm was distinctly smooth to sometimes granular and pale pink, with a “ground-glass” appearance. Nuclei were enlarged, and nucleoli were prominent and centrally located. A classical example is illustrated in Fig. 10. For the eosinophilic foci evaluated (n=11), 10/11 (91%) were in male rats.

Basophilic foci: Basophilic foci may show some resemblance with small cell change as observed in humans. Different subtypes have been described²⁶. The tigroid subtype consists of a focus of (small) basophilic cells distinct from the surrounding liver parenchyma and arranged in tortuous cords. Cells display large abundant basophilic bodies often arranged in clumps or long bands with a striped (“tigroid”) pattern in the paranuclear or peripheral regions of the cytoplasm (due to increased rough endoplasmic reticulum). In the samples we evaluated, the diffuse subtype was most common and consisted of small, discrete, clearly demarcated, strongly basophilic foci with irregular borders. These foci were randomly distributed within the hepatic lobule. Although different subtypes have been recognized in rodents, a typical example is shown in Fig. 12. Of the basophilic foci evaluated (n=9), 8/9 (89%) were observed in male rats.

Clear cell foci: Clear cell areas of groups of hepatocytes can be observed in HCCs in both human and rodents^{40–43}. Clear cell foci consisted of normal or enlarged groups of cells with prominent cell membranes and distinct cytoplasmic clear spaces surrounding a densely stained centrally located nucleus. Some eosinophilic or basophilic cells were occasionally present within clear cell foci. A classical example of a clear cell focus is presented in Fig. 13.

The role of clear cell foci in hepatocarcinogenesis is elusive and poorly described, although metabolic changes in carbohydrate metabolism have been associated with HCCs in both humans and rodents^{41,42,44,45}, and therefore these foci, as observed in the rat liver, could play a role in liver tumor formation. The selected clear cell foci (n=9) were only found in the livers of male rats.

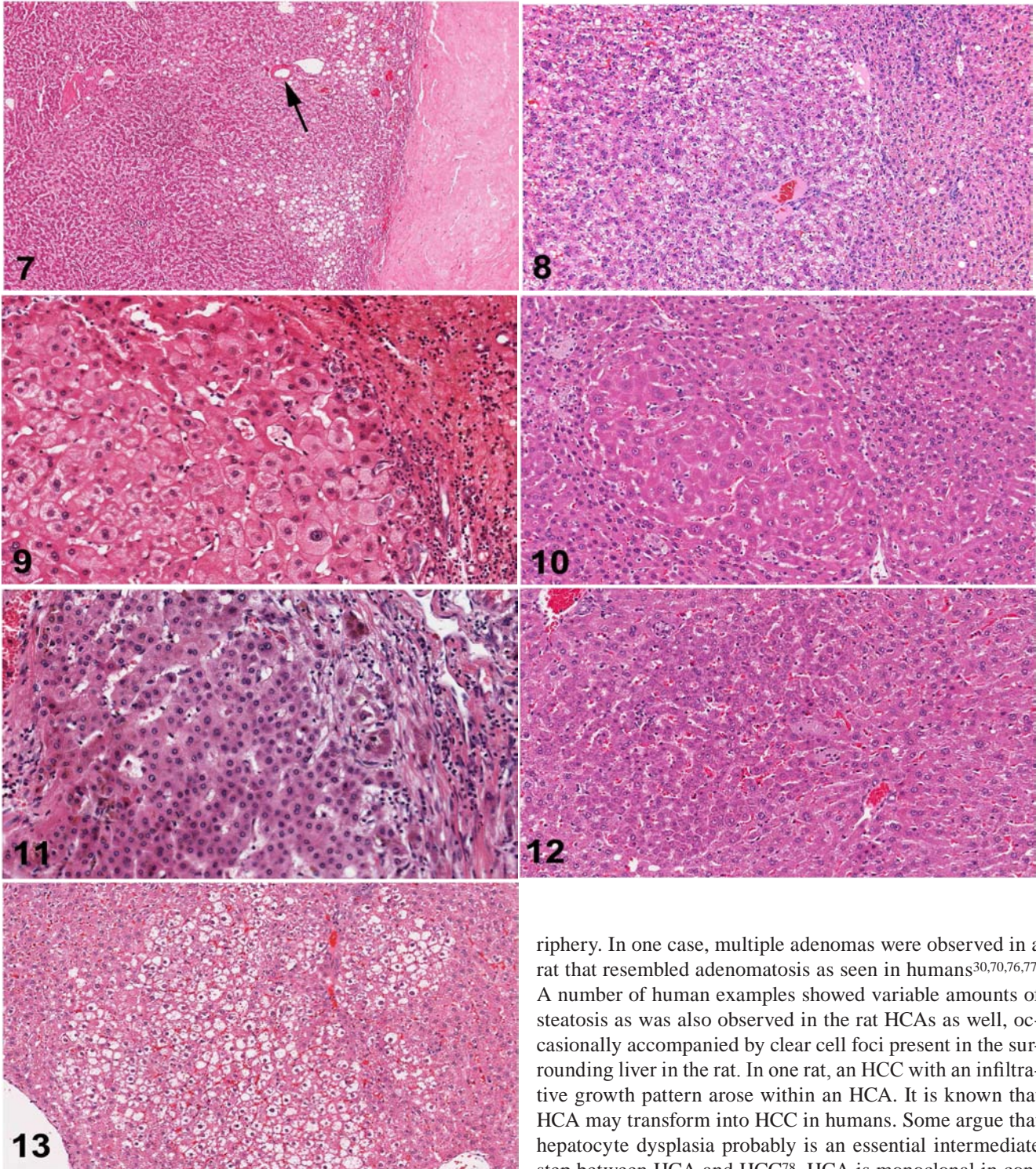
Discussion and Conclusion

The liver is a major target organ in preclinical toxicity and oncogenicity safety assessment studies with rodents. The significance of hepatic neoplastic findings in animal models has been questioned with regard to their predictive

value, as humans appear resistant to many agents that readily produce liver tumors in rodents⁴⁶. As toxicity to the liver is reported to be the second most frequent cause of drug failure due to adverse effects in clinical trials of potential drugs^{15,47,48}, the early detection and interpretation of proliferative lesions as well as nonproliferative hepatic lesions is of vital importance. One of the safety issues after long-term administration of a xenobiotic is carcinogenicity assessment, and both early and late proliferative liver lesions might be indicative of potential hepatocarcinogenesis or carcinogenesis at other sites in humans^{1,16,37,49–54}. The human and rat cases selected for this review were all morphologically consistent with descriptive features in texts and the peer reviewed literature using the most common and contemporary diagnostic criteria for both human and rat hepatic lesions^{8,22–24,26}.

We have reviewed FCAs (eosinophilic, basophilic and clear cell foci) and compared them with the human counterparts of LCC and SCC. LCC and SCC in humans and eosinophilic and basophilic FCA in the rat showed common histomorphological characteristics (Figs. 9 to 12), which might be indicative of a mutual presumptive role in the process of hepatocarcinogenesis. FNH is the second most common benign lesion of the liver in humans and occurs more often in young females but can occur in both sexes of adults^{55–60}. Cases in children have been reported^{61–63}. FNH is a benign lesion, and in contrast to HCA, the risk of complications such as hemorrhage and malignant transformation is virtually absent. The more common occurrence of FNH among young women is in line with our results that show that nearly all FNHs selected for this study were obtained from women (91.7%). FNH is a rare lesion in animals^{64,65}. Immunohistochemistry showed characteristic mild and focal cytokeratin 7 staining of hepatocytes, whereas cytokeratin 7 and cytokeratin 19 show a strong staining of bile ductules in the fibrous septa³⁹. The majority of patients with FNH have normal liver test results and alpha-fetoprotein is mostly in the normal range⁶⁶. Diagnosis of FNH may be difficult in humans if one or more major features are not prominent (central scar, ductular reaction) or if the FNH is steatotic, or has small nodules⁵⁷. A clear overview of the clinical and morphological features of FNH is presented by Ferrell and Kakar²⁴ for distinction between FNH, hepatocellular adenoma and HCC. Immunohistochemistry has also been extensively investigated and can be supportive for the differential diagnosis^{10,28,39,67}. The current opinion regarding the etiology of this lesion is that it represents a hyperplastic and altered growth of hepatocytes surrounding a pre-existing arterial malformation in response to changes in blood flow in the parenchyma^{8,23,68}. Treatment of symptomatic FNH in humans consists first of embolization and then resection⁶⁹.

Following hepatic angioma and FNH, HCA is the third most common benign proliferative lesion in humans and is known to occur in 85% of young female patients taking oral contraceptives^{70,71}. There is considerable overlap in morphologic features of well-differentiated hepatocellular lesions, necessitating the use of immunohistochemistry and other



techniques for diagnosis^{39,72-75}. More current diagnostic criteria distinguish four different subtypes of human HCAs based on their histological and molecular characteristics²⁹. The human HCAs examined were all from female patients, in line with the epidemiology and common occurrence of these tumors. HCA histological features observed in the rat were similar to those in humans. The tumors showed focal fatty change/steatosis and increased numbers of mitoses and there was also normal pre-existing liver present at the pe-

riphery. In one case, multiple adenomas were observed in a rat that resembled adenomatosis as seen in humans^{30,70,76,77}. A number of human examples showed variable amounts of steatosis as was also observed in the rat HCAs as well, occasionally accompanied by clear cell foci present in the surrounding liver in the rat. In one rat, an HCC with an infiltrative growth pattern arose within an HCA. It is known that HCA may transform into HCC in humans. Some argue that hepatocyte dysplasia probably is an essential intermediate step between HCA and HCC⁷⁸. HCA is monoclonal in contrast to FNH (polyclonal) and consequently has an inherent risk for progression to HCC⁷⁸. Moreover, β -catenin-mutated HCA has an increased risk of undergoing malignant change⁷⁹. Both the human and rat hepatocellular carcinomas reviewed in this study had malignant growths pattern that are typical for these tumors. Microscopically, the WHO distinguishes trabecular, acinar (pseudoglandular), scirrhous and solid forms⁸. Special histological subtypes, not included in this review, are the clear cell, fibrolamellar and mixed hepatocholangiocellular variants. Trabecular growth pat-

terns were the most common in both human and rat HCCs (Figs. 3 and 4) evaluated; sometimes mixtures of trabecular, solid and acinar/pseudoglandular patterns (Figs. 5 and 6) were also present in our collection. Both human and rat HCCs showed common histopathological characteristics that are typical for these malignant liver lesions. Liver cell dysplasia (LCD) is described in liver transplants containing underlying liver disease. These dysplastic hepatocytes can frequently be observed in the cirrhotic liver^{19,80,81} and have been proposed to contain precancerous properties^{31,38,82}. The cytological criteria for the diagnosis of LCD include cytoplasmic and nuclear changes, nuclear crowding or pleomorphism together with prominent nucleoli, hyperchromasia and sometimes multinucleation. The cytological features of liver cell dysplasia can strikingly mimic HCC⁸³ suggesting it is a putative preneoplastic lesion that can precede HCC in various species^{1,10,41,45,84–87}. The precancerous nature of both LCC and SCC with regard to progression to HCC is somewhat controversial, but some claim that either one or both of them have been associated with development of HCC^{31,35,38,49,82,87–90}.

In rats, FCAs have likewise been designated to play a precursor role in the process of hepatocarcinogenesis as they represent a localized proliferation of hepatocytes that are phenotypically different from the surrounding liver. These FCAs occur spontaneously in aged rats and other rodents and can be induced by chemical treatment. The incidence of spontaneous foci is highest in rats and can reach nearly 100% in F344 rats by the age of 2 years⁹¹. After administration of hepatocarcinogens, their incidence, size and/or multiplicity are usually increased, and latency usually decreased^{25,41,86}. These foci have been described in a number of animal models and are considered as precursor lesions of HCC⁴⁵, but controversy still remains. Some of these foci may have autonomous growth potential and may show enzyme profiles different from the normal hepatocytes (e.g., positive for γ -glutamyl transpeptidase, α -fetoprotein, glutathione S-transferase placental form, and negative for glucose-6-phosphatase and glycogen phosphorylase). However, it has been shown that certain conditions are required for promotion and progression of initiated cells. In addition, different mechanisms of promotion by different chemicals have been demonstrated in the multistep process of carcinogenesis, and it is stated that not all foci become

neoplasms^{91–94}. Some foci of cellular alteration can even regress, and different types of foci have different potentials for developing into neoplasms^{17,95}.

Since controversy with regard to the significance of presumptive preneoplastic liver lesions still exists, comparative research of these proliferative lesions in both rats and humans is needed.

Based on comparison of the histomorphological features of common nonneoplastic and neoplastic hepatocyte lesions of rats and humans, it is apparent that there are major similarities in diagnostic features, growth patterns, and behavior of these lesions in both species. Further study of presumptive preneoplastic lesions should help to further define their role in progression to malignancy and to provide a basis for using liver responses in rodents exposed to xenobiotics in safety assessment studies to predict potential risks to humans. Morphological similarities as illustrated in this review will be a first step to understanding their significance and relevance in human and animal liver tumor formation.

Acknowledgements: The authors thank the following people for their technical histology and editorial support: Ronald Herbert (NIEHS/NTP Archives), Keith Connelly (EPL, Inc.), Emily Singletary (EPL, Inc.), Mary Ellen Sutphin (EPL, Inc.) and Suzy Tirtodikromo (Global Pathology Support). Special thanks to Nikolas Stathonikos (UMCU) for photography support.

The majority of the photomicrographs used in this document were provided courtesy of the Division of Laboratories and Pharmacy, Department of Pathology, University Medical Center Utrecht, and the National Toxicology Program Archives, National Institute of Environmental Health Sciences, Research Triangle Park, NC, USA.

This research was supported [in part] by the Division of the National Toxicology Program of the NIH, National Institute of Environmental Health Sciences. This article may be the work product of an employee or group of employees of the National Institute of Environmental Health Sciences (NIEHS), National Institutes of Health (NIH), however, the statements, opinions or conclusions contained therein do not necessarily represent the statements, opinions or conclusions of NIEHS, NIH or the United States government.

Fig. 7. Human liver. Hepatocellular adenoma with multifocal steatosis and thick-wall blood vessel (arrow). Prominent fibrosis is present on right edge of the figure. H&E.

Fig. 8. Rat liver. Hepatocellular adenoma. The adenoma is sharply demarcated with slight compression of the adjacent normal parenchyma. H&E.

Fig. 9. Human liver. A classic example of large cell change. H&E.

Fig. 10. Rat liver. Eosinophilic focus of cellular alteration consisting of enlarged hepatocytes with increased acidophilic staining compared to the surrounding hepatic parenchyma. H&E.

Fig. 11. Human liver. Small cell change comprised of a focus of small cells with an irregular margin within a cirrhotic liver. H&E.

Fig. 12. Rat liver. A basophilic focus of cellular alteration with irregular but discrete margins surrounded by more eosinophilic normal parenchymal hepatocytes. H&E.

Fig. 13. Rat liver. Clear cell focus of cellular alteration with an irregular border and comprised of hepatocytes with clear cytoplasm and a centrally located nucleus. H&E.

References

1. Aleem E, Nehrbass D, Klimek F, Mayer D, and Bannasch P. Upregulation of the insulin receptor and type I insulin-like growth factor receptor are early events in hepatocarcinogenesis. *Toxicol Pathol.* **39**: 524–543. 2011. [[Medline](#)] [[CrossRef](#)]
2. Lee JS, Heo J, Libbrecht L, Chu IS, Kaposi-Novak P, Calvisi DF, Mikaelyan A, Roberts LR, Demetris AJ, Sun Z, Nevens F, Roskams T, and Thorgeirsson SS. A novel prognostic subtype of human hepatocellular carcinoma derived from hepatic progenitor cells. *Nat Med.* **12**: 410–416. 2006. [[Medline](#)] [[CrossRef](#)]
3. Frau M, Biasi F, Feo F, and Pascale RM. Prognostic markers and putative therapeutic targets for hepatocellular carcinoma. *Mol Aspects Med.* **31**: 179–193. 2010. [[Medline](#)] [[CrossRef](#)]
4. Lu X, Guo H, Molter J, Miao H, Gerber L, Hu Y, Barnes EL, Vogel H, Lee Z, Luo G, and Wang B. Alpha-fetoprotein-thymidine kinase-luciferase knockin mice: A novel model for dual modality longitudinal imaging of tumorigenesis in liver. *J Hepatol.* **55**: 96–102. 2011. [[Medline](#)] [[CrossRef](#)]
5. Stefaniuk P, Cianciara J, and Wiercinska-Drapalo A. Present and future possibilities for early diagnosis of hepatocellular carcinoma. *World J Gastroenterol.* **16**: 418–424. 2010. [[Medline](#)] [[CrossRef](#)]
6. Lopez JB. Recent developments in the first detection of hepatocellular carcinoma. *Clin Biochem Rev.* **26**: 65–79. 2005. [[Medline](#)]
7. Badvie S. Hepatocellular carcinoma. *Postgrad Med J.* **76**: 4–11. 2000. [[Medline](#)] [[CrossRef](#)]
8. Bosman FT, Carneiro F, Hruban RH, and Theise ND. Tumours of the liver and intrahepatic bile ducts. In: WHO Classification of Tumours of the Digestive System, 4. International Agency for Research on Cancer, Lyon. 195–254. 2010.
9. Zender L, Villanueva A, Tovar V, Sia D, Chiang DY, and Llovet JM. Cancer gene discovery in hepatocellular carcinoma. *J Hepatol.* **52**: 921–929. 2010. [[Medline](#)] [[CrossRef](#)]
10. Borbath I, Leclercq IA, and Sempoux C. barca-Quinones J, Desaegeer C, and Horsmans Y. Efficacy of lanreotide in preventing the occurrence of chemically induced hepatocellular carcinoma in rats. *Chem-Biol Interactions.* **183**: 238–248. 2010. [[CrossRef](#)]
11. Ascha MS, Hanouneh IA, Lopez R, Tamimi TA, Feldstein AF, and Zein NN. The incidence and risk factors of hepatocellular carcinoma in patients with nonalcoholic steatohepatitis. *Hepatology (Baltimore, Md).* **51**: 1972–1978. 2010. [[Medline](#)] [[CrossRef](#)]
12. Sanyal AJ, Yoon SK, and Lencioni R. The etiology of hepatocellular carcinoma and consequences for treatment. *The oncologist.* **15**: 14–22. 2010. [[Medline](#)] [[CrossRef](#)]
13. Liu Y, and Wu F. Global Burden of Aflatoxin-Induced Hepatocellular Carcinoma: A Risk Assessment. *Environ Health Perspect.* **118**: 818–824. 2010. [[Medline](#)] [[CrossRef](#)]
14. Spangenberg HC, Thimme R, and Blum HE. Serum markers of hepatocellular carcinoma. *Semin Liver Dis.* **26**: 385–390. 2006. [[Medline](#)] [[CrossRef](#)]
15. Tsuda H, Futakuchi M, Fukamachi K, Shirai T, Imaida K, Fukushima S, Tatematsu M, Furukawa F, Tamano S, and Ito N. A medium-term, rapid rat bioassay model for the detection of carcinogenic potential of chemicals. *Toxicol Pathol.* **38**: 182–187. 2010. [[Medline](#)] [[CrossRef](#)]
16. Bannasch P, Haertel T, and Qin S. Significance of Hepatic Preneoplasia in Risk Identification and Early Detection of Neoplasia. *Toxicol Pathol.* **31**: 134–139. 2003. [[Medline](#)]
17. Williams GM. The significance of chemically-induced hepatocellular altered foci in rat liver and application to carcinogen detection. *Toxicol Pathol.* **17**: 663–672. 1989. [[Medline](#)]
18. Terada T, and Nakanuma Y. Multiple occurrence of borderline hepatocellular nodules in human cirrhotic livers: possible multicentric origin of hepatocellular carcinoma. *Virchows Arch.* **427**: 379–383. 1995. [[Medline](#)] [[CrossRef](#)]
19. Cohen C, and Berson SD. Liver cell dysplasia in normal, cirrhotic, and hepatocellular carcinoma patients. *Cancer.* **57**: 1535–1538. 1986. [[Medline](#)] [[CrossRef](#)]
20. Cogliati B, Aloia TP, Bosch RV, Alves VA, Hernandez-Blazquez FJ, and Dagli ML. Identification of hepatic stem/progenitor cells in canine hepatocellular and cholangiocellular carcinoma. *Vet Comp Oncol.* **8**: 112–121. 2010. [[Medline](#)] [[CrossRef](#)]
21. Park YN. Update on precursor and early lesions of hepatocellular carcinomas. *Arch Pathol Lab Med.* **135**: 704–715. 2011. [[Medline](#)]
22. Goodman ZD, Ishak KG, Ferrell LD, and Geisinger KR. Hepatobiliary system and pancreas. In: *Surgical Pathology and Cytopathology*, 4th. SG Silverberg, RA DeLellis, WJ Frable, VA Livolsi, and MR Wick (eds). Elsevier, Philadelphia. 1465–1547. 2006.
23. Ishak KG, Goodman ZD, and Stocker JT. Tumors of the Liver and Intrahepatic Bile Ducts. Third series, Fascicle 31. Armed Forces Institute of Pathology, Washington, D.C. 2001.
24. Ferrell L, and Kakar S. Tumors of the liver, gallbladder, and biliary tree. In: *Diagnostic Histopathology of Tumors*, 3rd. CDM Fletcher (ed). Churchill Livingstone, Elsevier, Edinburgh. 417–461. 2007.
25. Bannasch P, and Zerban H. Tumours of the liver. In: *Pathology of tumours in laboratory animals. Tumours of the rat*, vol 1, 2nd. Turusov V and Mohr, U (eds) IARC scientific publications. 199–240. 1990. [[Medline](#)]
26. Thoolen B, Maronpot R, Harada T, Nyska A, Rousseaux C, Nolte T, Malarkey D, Kaufmann W, Küttler K, Deschl U, Nakae D, Gregson R, Vinlove M, Brix A, Singh B, Belpoggi F, and Ward J. Proliferative and Nonproliferative Lesions of the Rat and Mouse Hepatobiliary System. *Toxicol Pathol.* **38**: 5S–81S. 2010. [[Medline](#)] [[CrossRef](#)]
27. Roskams T, De VR, and Desmet V. ‘Undifferentiated progenitor cells’ in focal nodular hyperplasia of the liver. *Histopathology.* **28**: 291–299. 1996. [[Medline](#)] [[CrossRef](#)]
28. van Aalten SM, Verheij J, Terkivatan T, Dwarkasing RS, de Man RA, and IJzermans JNM. Validation of a liver adenoma classification system in a tertiary referral centre: Implications for clinical practice. *J Hepatol.* **55**: 120–125. 2011. [[Medline](#)] [[CrossRef](#)]
29. Bioulac-Sage P, Rebouissou S, Thomas C, Blanc JF, Saric J, Sa CA, Rullier A, Cubel G, Couchy G, Imbeaud S, Balabaud C, and Zucman-Rossi J. Hepatocellular adenoma subtype classification using molecular markers and immunohistochemistry. *Hepatology (Baltimore, Md).* **46**: 740–748. 2007. [[Medline](#)] [[CrossRef](#)]
30. Rebouissou S, Imbeaud S, Jeannot E, Saric J, Balabaud C, Bioulac-Sage P, and Zucman-Rossi J. Role of HNF1 inac-

- tivation in hepatocellular adenoma lipogenesis. *J Hepatol.* **44**: S44. 2006. [[CrossRef](#)]
31. Anthony PP, Vogel CL, and Barker LF. Liver cell dysplasia: a premalignant condition. *J Clin Pathol.* **26**: 217–223. 1973. [[Medline](#)] [[CrossRef](#)]
 32. An CST, Petrovic LM, Reyter I, Tolmachoff T, Ferrell LD, Thung SN, Geller SA, and Marchevsky AM. The application of image analysis and neural network technology to the study of large-cell liver-cell dysplasia and hepatocellular carcinoma. *Hepatology (Baltimore, Md).* **26**: 1224–1231. 1997. [[Medline](#)] [[CrossRef](#)]
 33. Ganne-Carrié N, Chastang C, Chapel F, Munz C, Pateron D, Sibony M, Deny P, Trinchet JC, Callard P, Guettier C, and Beaugrand M. Predictive score for the development of hepatocellular carcinoma and additional value of liver large cell dysplasia in Western patients with cirrhosis. *Hepatology (Baltimore, Md).* **23**: 1112–1118. 1996. [[Medline](#)] [[CrossRef](#)]
 34. Ikeda H, Sasaki M, Sato Y, Harada K, Zen Y, Mitsui T, and Nakanuma Y. Large cell change of hepatocytes in chronic viral hepatitis represents a senescent-related lesion. *Hum Pathol.* **40**: 1774–1782. 2009. [[Medline](#)] [[CrossRef](#)]
 35. Le Bail B, Bernard PH, Carles J, Balabaud C, and Bioulac-Sage P. Prevalence of liver cell dysplasia and association with HCC in a series of 100 cirrhotic liver explants. *J Hepatol.* **27**: 835–842. 1997. [[Medline](#)] [[CrossRef](#)]
 36. Lee RG, Tsamandas AC, and Demetris AJ. Large cell change (liver cell dysplasia) and hepatocellular carcinoma in cirrhosis: matched case-control study, pathological analysis, and pathogenetic hypothesis. *Hepatology (Baltimore, Md).* **26**: 1415–1422. 1997. [[Medline](#)] [[CrossRef](#)]
 37. Adachi E, Hashimoto H, and Tsuneyoshi M. Proliferating cell nuclear antigen in hepatocellular carcinoma and small cell liver dysplasia. *Cancer.* **72**: 2902–2909. 1993. [[Medline](#)] [[CrossRef](#)]
 38. Watanabe S, Okita K, Harada T, Kodama T, Numa Y, Takemoto T, and Takahashi T. Morphologic studies of the liver cell dysplasia. *Cancer.* **51**: 2197–2205. 1983. [[Medline](#)] [[CrossRef](#)]
 39. Ahmad I, Iyer A, Marginean CE, Yeh MM, Ferrell L, Qin L, Bifulco CB, and Jain D. Diagnostic use of cytokeratins, CD34, and neuronal cell adhesion molecule staining in focal nodular hyperplasia and hepatic adenoma. *Hum Pathol.* **40**: 726–734. 2009. [[Medline](#)] [[CrossRef](#)]
 40. Maronpot RR, Montgomery CA Jr, Boorman GA, and McConnell EE. National toxicology program nomenclature for hepatoproliferative lesions of rats. *Toxicol Pathol.* **14**: 263–273. 1986. [[Medline](#)] [[CrossRef](#)]
 41. Bannasch P, Enzmann H, Klimek F, Weber E, and Zerban H. Significance of sequential cellular changes inside and outside foci of altered hepatocytes during hepatocarcinogenesis. *Toxicol Pathol.* **17**: 617–628. 1989. [[Medline](#)]
 42. Bannasch P. Hepatocellular glycogenesis and hepatic neoplasms. *Toxicol Pathol.* **38**: 1000–1002. 2010. [[Medline](#)] [[CrossRef](#)]
 43. Enzmann H, Kaliner G, Watta-Gebert B, and Löser E. Foci of altered hepatocytes induced in embryonal turkey liver. *Carcinogenesis.* **13**: 943–946. 1992. [[Medline](#)] [[CrossRef](#)]
 44. Enzmann H, Zerban H, Löser E, and Bannasch P. Glycogen phosphorylase hyperactive foci of altered hepatocytes in aged rats. *Virchows Arch B.* **62**: 3–8. 1992. [[Medline](#)] [[CrossRef](#)]
 45. Libbrecht L, Desmet V, and Roskams T. Review Article: Preneoplastic lesions in human hepatocarcinogenesis. *Liver Int.* **25**: 16–27. 2005. [[Medline](#)] [[CrossRef](#)]
 46. Anthony PP. Liver tumours. *Baillière's Clin Gastroenterol.* **2**: 501–522. 1988. [[Medline](#)] [[CrossRef](#)]
 47. Foster JR. Spontaneous and drug-induced hepatic pathology of the laboratory Beagle dog, the cynomolgus macaque and the marmoset. *Toxicol Pathol.* **33**: 63–74. 2005. [[Medline](#)] [[CrossRef](#)]
 48. Lee S, Lee HJ, Kim JH, Lee HS, Jang JJ, and Kang GH. Aberrant CpG island hypermethylation along multistep hepatocarcinogenesis. *Am J Pathol.* **163**: 1371–1378. 2003. [[Medline](#)] [[CrossRef](#)]
 49. Cheah PL, Looi LM, Nazarina AR, Goh KL, Rosmawati M, and Vijeyasingam R. Histopathological landmarks of hepatocellular carcinoma in Malaysians. *Malays J Pathol.* **25**: 37–43. 2003. [[Medline](#)]
 50. Hoenerhoff MJ, Pandiri AR, Lahousse SA, Hong H-H, Ton T-V, Masinde T, Sills RC, Auerbach SS, Gerrish K, Bushel PR, Shockley KR, and Peddada SD. Global gene profiling of spontaneous hepatocellular carcinoma in B6C3F1 Mice: Similarities in the molecular landscape with human liver cancer. *Toxicol Pathol.* **39**: 678–699. 2011. [[Medline](#)] [[CrossRef](#)]
 51. Andersen JB, Loi R, Perra A, Factor VM, Ledda-Columbano GM, Columbano A, and Thorgerirsson SS. Progenitor-derived hepatocellular carcinoma model in the rat. *Hepatology (Baltimore, Md).* **51**: 1401–1409. 2010. [[Medline](#)] [[CrossRef](#)]
 52. Cohen SM. An enhanced 13-week bioassay: An alternative to the 2-year bioassay to screen for human carcinogenesis. *Exp Toxicol Pathol.* **62**: 497–502. 2010. [[Medline](#)] [[CrossRef](#)]
 53. Marquardt JU, and Thorgerirsson SS. Stem cells in hepatocarcinogenesis: evidence from genomic data. *Semin Liver Dis.* **30**: 26–34. 2010. [[Medline](#)] [[CrossRef](#)]
 54. Samson TJ. Role of cancer stem cells in hepatocarcinogenesis. *Gen Med.* **3**: 11. 2011.
 55. Lefkowitz JH. Advances in hepatobiliary pathology: update for 2010. *Clin Liver Dis.* **14**: 747–762. 2010. [[Medline](#)] [[CrossRef](#)]
 56. Bellamy COC. Pathology of liver tumours. *Surgery (Oxford).* **28**: 183–188. 2010. [[CrossRef](#)]
 57. Bioulac-Sage P, Laumonier H, Rullier A, Cubel G, Laurent C, Zucman-Rossi J, and Balabaud C. Over-expression of glutamine synthetase in focal nodular hyperplasia: a novel easy diagnostic tool in surgical pathology. *Liver Int.* **29**: 459–465. 2009. [[Medline](#)] [[CrossRef](#)]
 58. Paradis V. Benign liver tumors: an update. *Clin Liver Dis.* **14**: 719–729. 2010. [[Medline](#)] [[CrossRef](#)]
 59. Blanc JF, Jeannot E, Laurent C, Sa Cunha A, Rebuissou A, Lepreux S, Le Bail B, Troutte H, Saric J, Balabaud C, Zucman-Rossi J, and Bioulac-Sage P. 232 Focal nodular telangiectasia (FNT) of the liver: Pathological and molecular characterization. *J Hepatol.* **40**: 74. 2004. [[CrossRef](#)]
 60. Kayhan A, Venu N, Lakadamyali H, Jensen D, and Oto A. Multiple progressive focal nodular hyperplasia lesions of liver in a patient with hemosiderosis. *World J Radiol.* **2**: 405–409. 2010. [[Medline](#)] [[CrossRef](#)]
 61. Farruggia P, Alaggio R, Cardella F, Tropicia S, Trizzino A, Ferrara F, and D'Angelo P. Focal nodular hyperplasia of the liver: an unusual association with diabetes mellitus in

- a child and review of literature. *Ital J Pediatr.* **36**: 41. 2010. [[Medline](#)] [[CrossRef](#)]
62. Sugito K, Uekusa S, Kawashima H, Furuya T, Ohashi K, Inoue M, Ikeda T, Koshinaga T, Tomita R, Mugishima H, and Maebayashi T. The clinical course in pediatric solid tumor patients with focal nodular hyperplasia of the liver. *Int J Clin Oncol.* **16**: 482–487. 2011. [[Medline](#)] [[CrossRef](#)]
 63. Towbin AJ, Luo GG, Mo JQ, and Mo JQ. Focal nodular hyperplasia in children, adolescents, and young adults. *Pediatr Radiol.* **41**: 341–349. 2011. [[Medline](#)] [[CrossRef](#)]
 64. Fujishima J, Satake S, Furukawa T, Kurokawa C, Kodama R, Moriyama A, Sasaki Y, Kamimura Y, and Maeda H. Focal nodular hyperplasia in the livers of cynomolgus macaques (*Macaca fascicularis*). *J Toxicol Pathol.* **24**: 125–129. 2011. [[Medline](#)] [[CrossRef](#)]
 65. Sumiyoshi M, Sakanaka M, and Kimura Y. Chronic intake of a high-cholesterol diet resulted in hepatic steatosis, focal nodular hyperplasia and fibrosis in non-obese mice. *Brit J Nutr.* **103**: 378–385. 2010. [[Medline](#)] [[CrossRef](#)]
 66. Mneineh W, Farges O, Bedossa P, Belghiti J, and Paradis V. High serum level of alpha-fetoprotein in focal nodular hyperplasia of the liver. *Pathol Int.* **61**: 491–494. 2011. [[Medline](#)] [[CrossRef](#)]
 67. Zhu ZW, Friess H, Wang L, bou-Shady M, Zimmermann A, Lander AD, Korc M, Kleeff J, and Büchler MW. Enhanced glypican-3 expression differentiates the majority of hepatocellular carcinomas from benign hepatic disorders. *Gut.* **48**: 558–564. 2001. [[Medline](#)] [[CrossRef](#)]
 68. Wanless IR, Mawdsley C, and Adams R. On the pathogenesis of focal nodular hyperplasia of the liver. *Hepatology* (Baltimore, Md). **5**: 1194–1200. 1985. [[Medline](#)] [[CrossRef](#)]
 69. Arts CH, van HR, de Kort GA, and Moll FL. Inferior caval vein thrombosis owing to compression of focal nodular hyperplasia: surgical resection after shrinkage by hepatic artery embolization. *Vascular.* **18**: 53–58. 2010. [[Medline](#)] [[CrossRef](#)]
 70. Maillette De Buy Wenniger L, Terpstra V, and Beuers U. Focal nodular hyperplasia and hepatic adenoma: Epidemiology and pathology. *Dig Surg.* **27**: 24–31. 2010. [[Medline](#)] [[CrossRef](#)]
 71. Bioulac-Sage P, Laumonier H, Laurent C, Zucman-Rossi J, and Balabaud C. Hepatocellular adenoma: what is new in 2008. *Hepatol Int.* **2**: 316–321. 2008. [[Medline](#)] [[CrossRef](#)]
 72. Di CI, Pulvirenti E, Toro A, and Priolo GD. Adenoma or atypical hepatic focal nodular hyperplasia: role of preoperative imaging and laparoscopic treatment. *Surg Lapar Endos & Percut Techn.* **20**: e105–e109. 2010. [[CrossRef](#)]
 73. Bioulac-Sage P, Balabaud C, and Zucman-Rossi J. Focal nodular hyperplasia, hepatocellular adenomas: past, present, future. *Gastroenterol Clin Biol.* **34**: 355–358. 2010. [[Medline](#)] [[CrossRef](#)]
 74. Bioulac-Sage P, Cubel G, Balabaud C, and Zucman-Rossi J. Revisiting the pathology of resected benign hepatocellular nodules using new immunohistochemical markers. *Semin Liver Dis.* **31**: 91–103. 2011. [[Medline](#)] [[CrossRef](#)]
 75. Shafizadeh N, and Kakar S. Diagnosis of well-differentiated hepatocellular lesions: role of immunohistochemistry and other ancillary techniques. *Adv Anat Pathol.* **18**: 438–445. 2011. [[Medline](#)] [[CrossRef](#)]
 76. Bioulac-Sage P, Balabaud C, and Zucman-Rossi J. Subtype classification of hepatocellular adenoma. *Dig Surg.* **27**: 39–45. 2010. [[Medline](#)] [[CrossRef](#)]
 77. Reznik Y, Dao T, Coutant R, Chiche L, Jeannot E, Clauin S, Rousselot P, Fabre M, Oberti F, Fatome A, Zucman-Rossi J, and Bellanne-Chantelot C. Hepatocyte nuclear factor-1 alpha gene inactivation: cosegregation between liver adenomatosis and diabetes phenotypes in two maturity-onset diabetes of the young (MODY)3 families. *Clin Endocrinol Metab.* **89**: 1476–1480. 2004. [[Medline](#)] [[CrossRef](#)]
 78. Farges O, and Dokmak S. Malignant transformation of liver adenoma: an analysis of the literature. *Dig Surg.* **27**: 32–38. 2010. [[Medline](#)] [[CrossRef](#)]
 79. Zucman-Rossi J, Jeannot E, Nhieu JT, Scoazec JY, Guettier C, Rebouissou S, Bacq Y, Leteurtre E, Paradis V, Michalak S, Wendum D, Chiche L, Fabre M, Mellottee L, Laurent C, Partensky C, Castaing D, Zafrani ES, Laurent-Puig P, Balabaud C, and Bioulac-Sage P. Genotype-phenotype correlation in hepatocellular adenoma: new classification and relationship with HCC. *Hepatology* (Baltimore, Md). **43**: 515–524. 2006. [[Medline](#)] [[CrossRef](#)]
 80. Kobayashi M, Ikeda K, Hosaka T, Sezaki H, Someya T, Akuta N, Suzuki F, Suzuki Y, Saitoh S, Arase Y, and Kumada H. Dysplastic nodules frequently develop into hepatocellular carcinoma in patients with chronic viral hepatitis and cirrhosis. *Cancer.* **106**: 636–647. 2006. [[Medline](#)] [[CrossRef](#)]
 81. Lefkowitz JH, and Apfelbaum TF. Liver cell dysplasia and hepatocellular carcinoma in non-A, non-B hepatitis. *Arch Pathol Lab Med.* **111**: 170–173. 1987. [[Medline](#)]
 82. Libbrecht L, Craninx M, Nevens F, Desmet V, and Roskams T. Predictive value of liver cell dysplasia for development of hepatocellular carcinoma in patients with non-cirrhotic and cirrhotic chronic viral hepatitis. *Histopathology.* **39**: 66–73. 2001. [[Medline](#)] [[CrossRef](#)]
 83. Tao LC. Oral contraceptive-associated liver cell adenoma and hepatocellular carcinoma. Cytomorphology and mechanism of malignant transformation. *Cancer.* **68**: 341–347. 1991. [[Medline](#)] [[CrossRef](#)]
 84. Podda M, Roncalli M, Battezzati PM, Borzio M, Bruno S, Servida E, and Coggi G. Liver-cell dysplasia and hepatocellular carcinoma. *Ital J Gastroenterol.* **24**: 39–42. 1992. [[Medline](#)]
 85. Enzmann H, Kuhlem C, Kaliner G, Löser E, and Bannasch P. Rapid induction of preneoplastic liver foci in embryonal turkey liver by diethylnitrosamine. *Toxicol Pathol.* **23**: 560–569. 1995. [[Medline](#)] [[CrossRef](#)]
 86. Maronpot RR, Pitot HC, and Peraino C. Use of rat liver altered focus models for testing chemicals that have completed two-year carcinogenicity studies. *Toxicol Pathol.* **17**: 651–662. 1989. [[Medline](#)]
 87. Koo JS, Kim H, Park B, Ahn S, Han KH, Chon C, Park C, and Park Y. Predictive value of liver cell dysplasia for development of hepatocellular carcinoma in patients with chronic hepatitis B. *J Clin Gastroenterol.* **42**: 738–743. 2008. [[Medline](#)] [[CrossRef](#)]
 88. Borzio M, Bruno S, Roncalli M, and Mels GC. Liver cell dysplasia is a major risk factor for hepatocellular carcinoma in cirrhosis: a prospective study. *Gastroenterology.* **108**: 812. 1995. [[Medline](#)] [[CrossRef](#)]
 89. Park YN, and Roncalli M. Large liver cell dysplasia: a controversial entity. *J Hepatol.* **45**: 734–743. 2006. [[Medline](#)] [[CrossRef](#)]
 90. Rubin EM, DeRose PB, and Cohen C. Comparative image

- cytometric DNA ploidy of liver cell dysplasia and hepatocellular carcinoma. *Mod Pathol.* **7**: 677–680. 1994. [[Medline](#)]
91. Narama I, Imaida K, Iwata H, Nakae D, Nishikawa A, and Harada T. A review of nomenclature and diagnostic criteria for proliferative lesions in the liver of rats by a working group of the Japanese Society of Toxicologic Pathology. *J Toxicol Pathol.* **16**: 1–17. 2003. [[CrossRef](#)]
92. Popp JA, and Goldsworthy TL. Defining foci of cellular alteration in short-term and medium-term rat liver tumor models. *Toxicol Pathol.* **17**: 561–568. 1989. [[Medline](#)]
93. Harada T, Maronpot RR, Morris RW, Stitzel KA, and Boorman GA. Morphological and stereological characterization of hepatic foci of cellular alteration in control Fischer 344 rats. *Toxicol Pathol.* **17**: 579–593. 1989. [[Medline](#)]
94. Pitot HC, Campbell HA, Maronpot R, Bawa N, Rizvi TA, Xu YH, Sargent L, Dragan Y, and Pyron M. Critical parameters in the quantitation of the stages of initiation, promotion, and progression in one model of hepatocarcinogenesis in the rat. *Toxicol Pathol.* **17**: 594–611. 1989. [[Medline](#)]
95. Enzmann H, and Bannasch P. Non-persisting early foci of altered hepatocytes induced in rats by N-nitrosomorpholine. *J Cancer Res Clin Oncol.* **114**: 30–34. 1988. [[Medline](#)] [[CrossRef](#)]