

Rodent Liver Tumors: NCI/NTP Historic Perspective

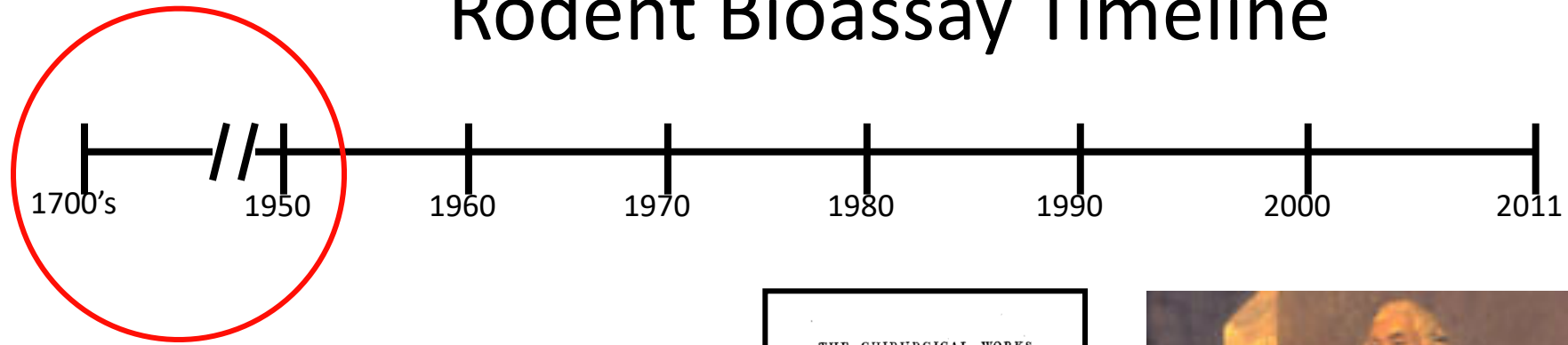
Bob Maronpot, Raleigh, NC

Rodent Liver Tumors: NCI/NTP Historic Perspective

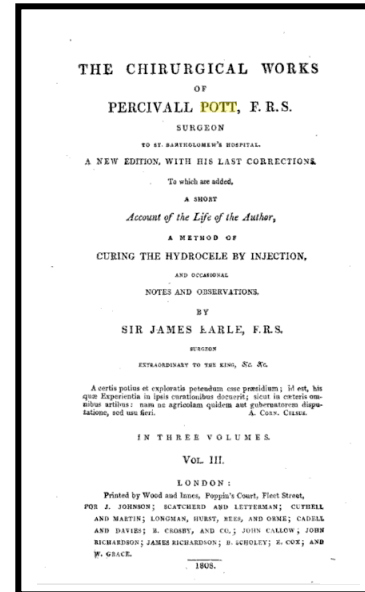
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- A little bit of NCI/NTP rodent bioassay history
- NTP liver tumor data
- Liver tumor images
- Current NTP safety assessment perspective

Rodent Bioassay Timeline

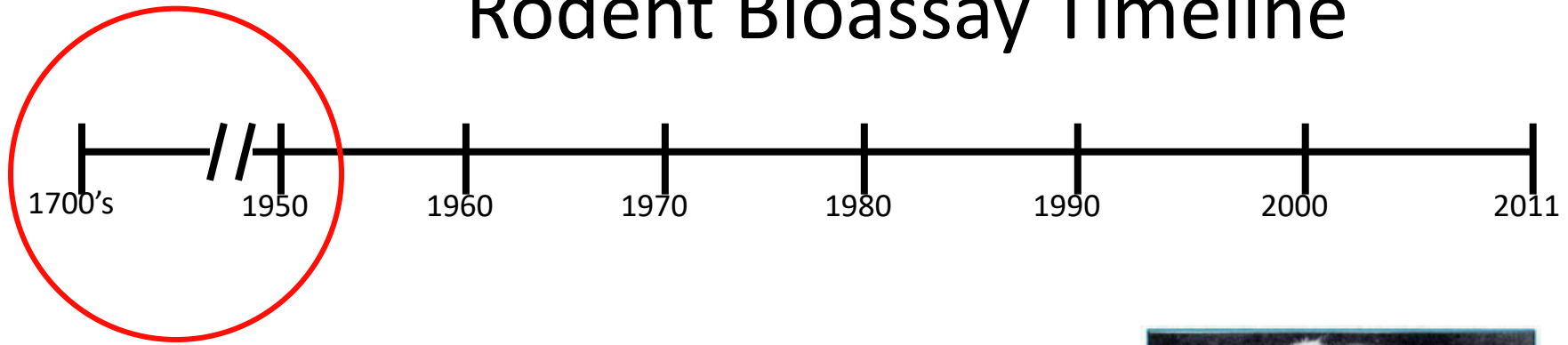


- Bernardino Ramazzini – 1713
 - Breast cancer in nuns
- John Hill – 1761
 - Snuff & oral/nasal cancer
- Percival Pott – 1775
 - Scrotal cancer
- Elmslie -1866 (Kashmir)
 - Epithelioma of abdomen & thighs
- Rehn – 1895
 - Bladder cancer in aniline dye workers
- Yamagiwa & Ichikawa – 1918
 - Tar & soot on rabbit ears



1775 - Scrotal cancer in chimney sweeps. Cancer was attributed to the tar and soot in the chimneys. This is an early and famous example of occupational cancer in humans.

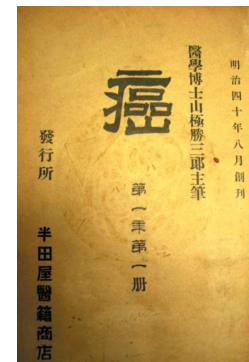
Rodent Bioassay Timeline



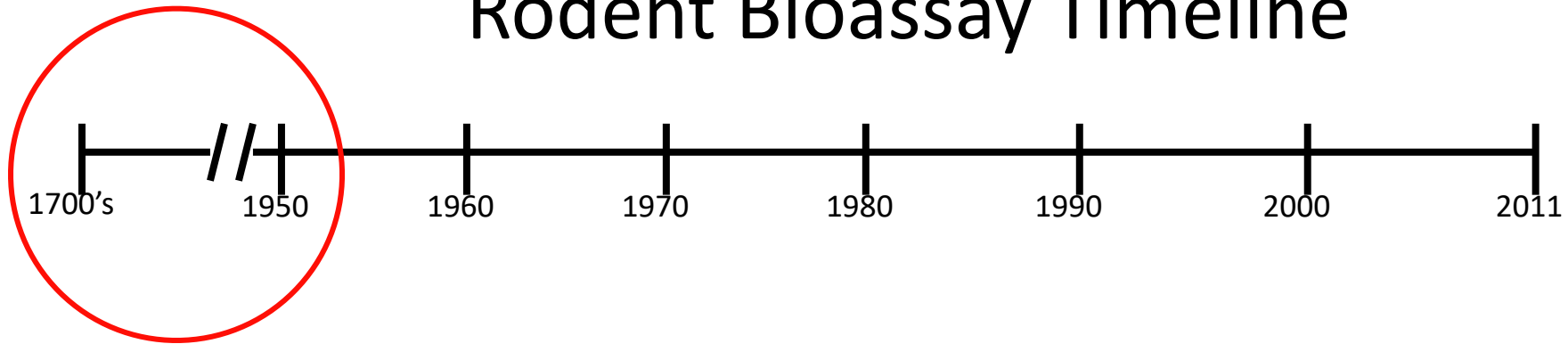
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**Katsusaburo Yamagiwa
(1863-1930)**

*“Cancer was produced!
Proudly I walk a few steps”*

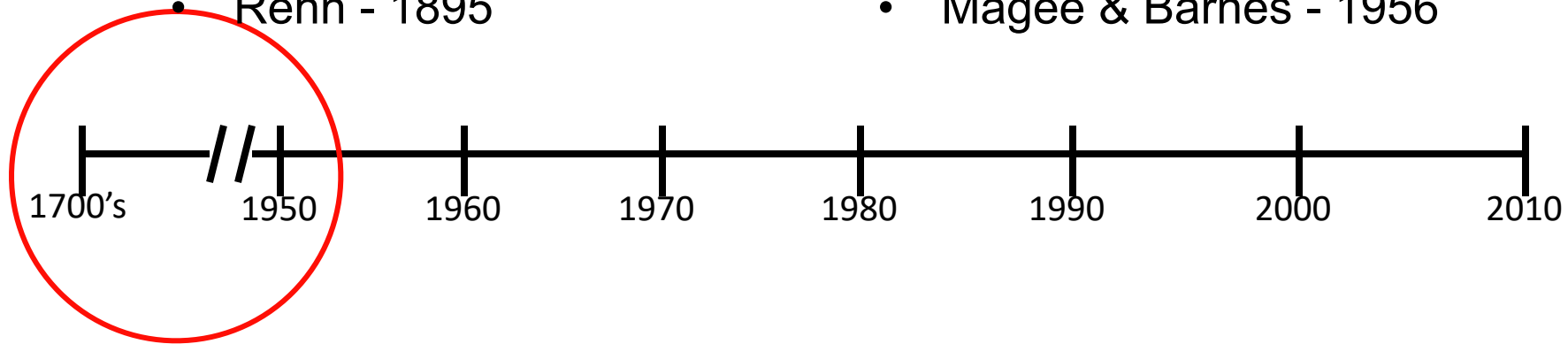


Rodent Bioassay Timeline



- Bernardino Ramazzini – 1713
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 - Bladder cancer in aniline dye workers
- Yamagiwa & Ichikawa – 1918
 - Tar & soot on rabbit ears
- Murphy & Sturm – 1925
 - Lung tumors in tar-painted mice
- Cook et al. – 1932
 - Cancer induction by PAHs
- Sasaki & Yoshida – 1935
 - o-amidoazotoluene diet and liver cancer; effects of dose on latency and use of stop studies
- Berenblum – 1941
 - Concept of co-carcinogenesis
 - Initiation, promotion, progression
- Magee & Barnes – 1956
 - Nitrosamines & liver cancer in rats

- Bernardino Ramazzini - 1713
- John Hill - 1761
- Percival Pott - 1775
- Elmslie - 1866
- Jonathon Hutchinson - 1888
- Rehn - 1895
- Yamagiwa & Ichikawa - 1918
- Murphy & Sturm - 1925
- Cook et al. - 1932
- Sasaki & Yoshida - 1935
- Berenblum - 1941
- Magee & Barnes - 1956



- Realization that chemicals, environmental factors, and aspects of lifestyle cause cancer

Concept of the rodent bioassay & its establishment by the National Cancer Institute (NCI)

- 1962 - First contracted bioassay
- 1969 - Innes et al*., study published
 - 20,000 mice; 127 different chemicals; 18-mo studies
 - Selection of B6C3F1 mouse
- 1971 - National Cancer Act
 - Decision made to standardize bioassay testing
- ~1975 – Inbred F344 rat selected
 - Small size, vigor & survival, disease resistance

* Innes et al., JNCI 42(6): 1104-1114 (1969)

Thou shalt use standardize tests



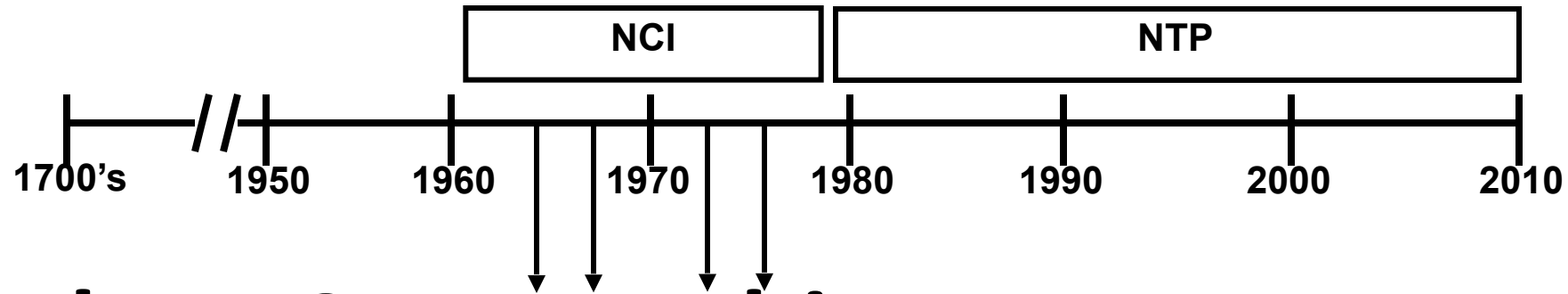
Thou shalt use two species



Thou shalt use the MTD & 1/2 MTD



CANCER BIOASSAY TIMELINE



The NCI cancer bioassay

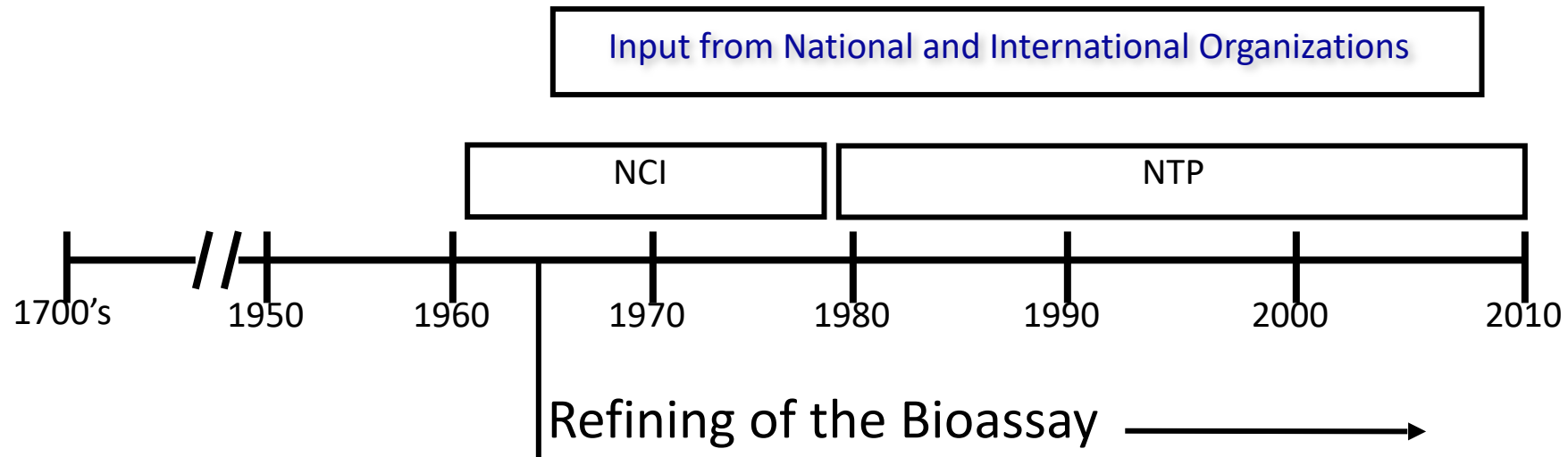
50 Male and 50 female **F344** rats

50 Male and 50 female **B6C3F1** mice

Maximum tolerated dose & 1/2 MTD

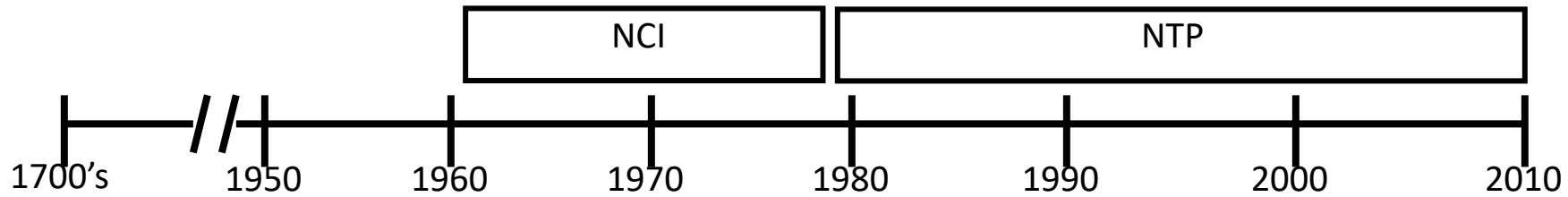
Test duration of 18 months or 2 years

Pathology evaluation



- Standardization of bioassay
 - Originally designed for screening
- Extensive pathology with peer review*
 - Standardization of diagnostic nomenclature
- Statistical evaluation standardized
- Historical control database
- Search for alternative models

*Maronpot & Boorman (1982) Toxicol Pathol 10(2): 71-78



Limitations of the bioassay

- Resource intensive
- Not validated
- Inherent insensitivity for detecting weak or moderate carcinogens
- Single chemical exposure vs “real world”
- Not sure if an agent has carcinogenic potential under actual human exposure conditions
- Debate regarding relevance
 - Rodent-specific mechanisms
 - High doses

Alternative models & ancillary approaches

- Strain A mouse
- Two-stage & neonatal models
- Humanized mice
- Ito medium-term model
- Transgenic models
- Local subcutaneous injection
- Medaka & guppy models
- Genotoxicity batteries

Target organs of chemical-induced carcinogenicity

#	Mouse (%), n=490*	Rat (%), n=490*
1	Liver (27.1)	Liver (10.6)
2	Lung (8.8)	Kidney, tubular cell (9.2)
3	Forestomach (4.7)	Mammary gland (5.9)
4	Hematopoietic system (4.5)	Lung (4.6)
5	Harderian gland Thyroid gland, follicular cell (2.7)	Thyroid gland, follicular cell (4.5)
6	Kidney, tubular cell (2.5)	Forestomach (4.3)
7	Vascular System (Unspecified) (2.3)	Urinary bladder (4.1)
8	Mammary gland (2.2)	Skin (3.8)
9	Ovary (2)	Hematopoietic system (3.7)
10	Skin (1.6)	Adrenal medulla Oral cavity Zymbal gland (3.5)

*n=490 studies where the same chemical was tested in both F344 rats and B6C3F1 mice

Background liver tumor incidence

Historical control incidences of liver tumors in rats (F344/N) and mice (B6C3F1)

Tumor type	Male Mouse % (Range%)	Female Mouse % (Range%)	Male Rat % (Range%)	Female Rat % (Range%)
Hepatocellular Adenoma	54.91 (34-78)	25.68 (10-67)	1.43 (0-6)	0.86 (0-4)
Hepatocellular Carcinoma	30 (16-50)	12.93 (4-20)	0.57 (0-4)	0.14 (0-2)
Hepatoblastoma	3.27 (0-8)	0.55 (0-2)	0	0
Combined	71.82 (62-84)	34.43 (16-73)	2 (0-6)	1 (0-4)

Mouse, n=550; Rat, n=700

Courtesy of A. Pandiri 2020

Frequency of tissue response in 290 cancer-positive NTP mouse and/or rat bioassays

<i>Liver</i>	<i>57 %</i>
<i>Lung</i>	<i>22 %</i>
<i>Kidney</i>	<i>22 %</i>
<i>Mammary gland</i>	<i>14 %</i>
<i>Hematopoietic</i>	<i>13 %</i>
<i>Forestomach</i>	<i>12 %</i>
<i>Thyroid</i>	<i>10 %</i>
<i>Vascular System</i>	<i>9 %</i>

Liver tumor incidences based on 490 studies

- 30% (146) of 490 NTP studies had an hepatocellular tumor response in rats and/or mice*
- Species dependence: mouse – 95/146 (65%), rat – 14/146 (9.6%), or both species 37/146 (25.3%)

Liver tumors N=146/490*	Mouse Male n (%)	Mouse Female n (%)	Rat Male n (%)	Rat Female n (%)
Nodule	0	1 (0.6)	10 (6.8)	6 (4.1)
Hepatocellular Adenoma	6 (4.1)	14 (9.5)	1 (0.6)	5 (3.4)
Hepatocellular carcinoma	64 (43.8)	88 (60.2)	31 (21.2)	30 (20.5)
Hepatoblastoma	20 (13.7)	13 (8.9)	0 (0)	1 (0.6)
Combined	90 (61.6)	116 (79.5)	42 (28.8)	42 (28.8)

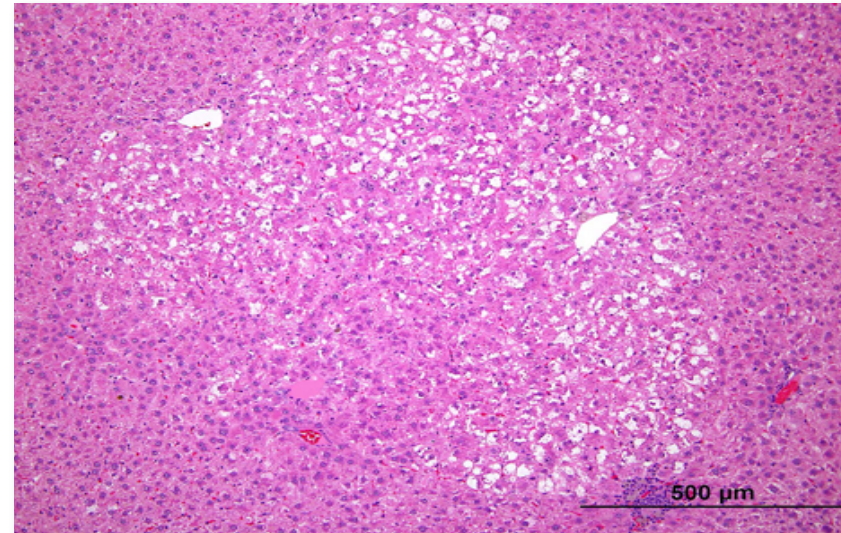
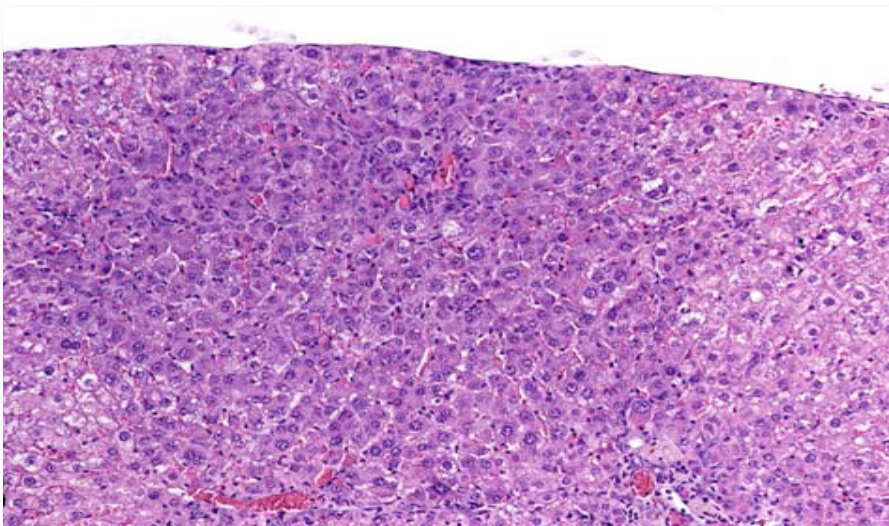
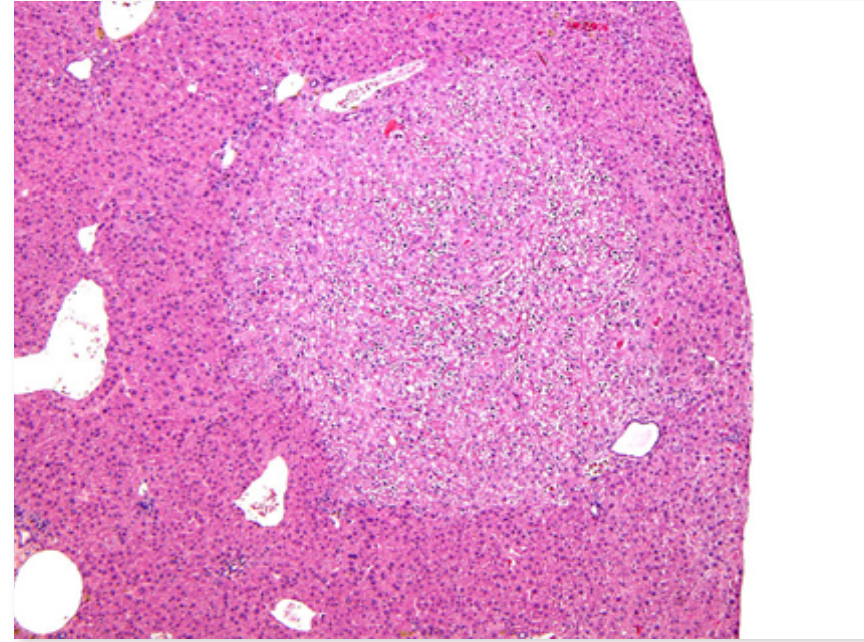
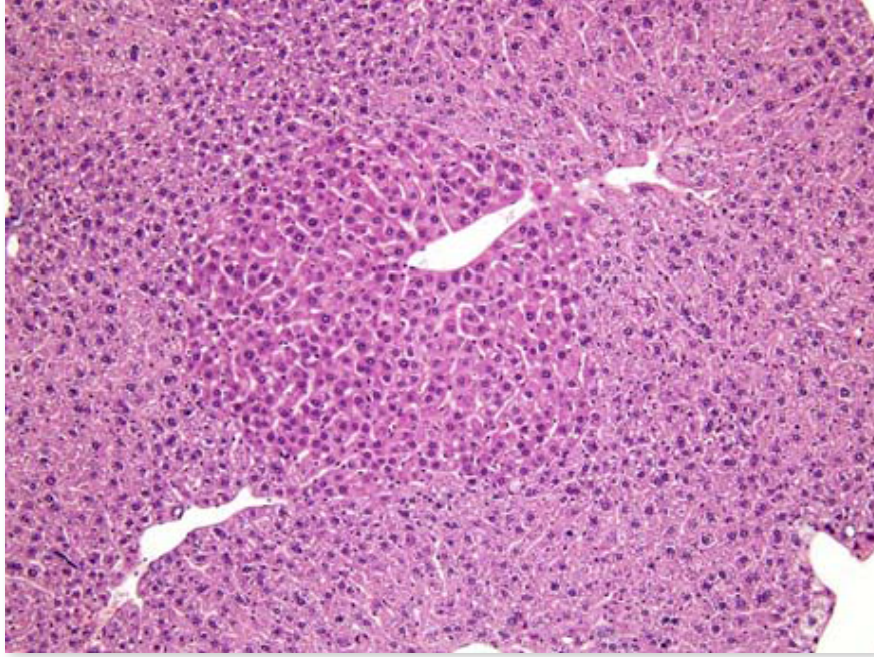
* 490 studies with same chemical tested in both rats and mice

Data courtesy of A. Pandiri 2022

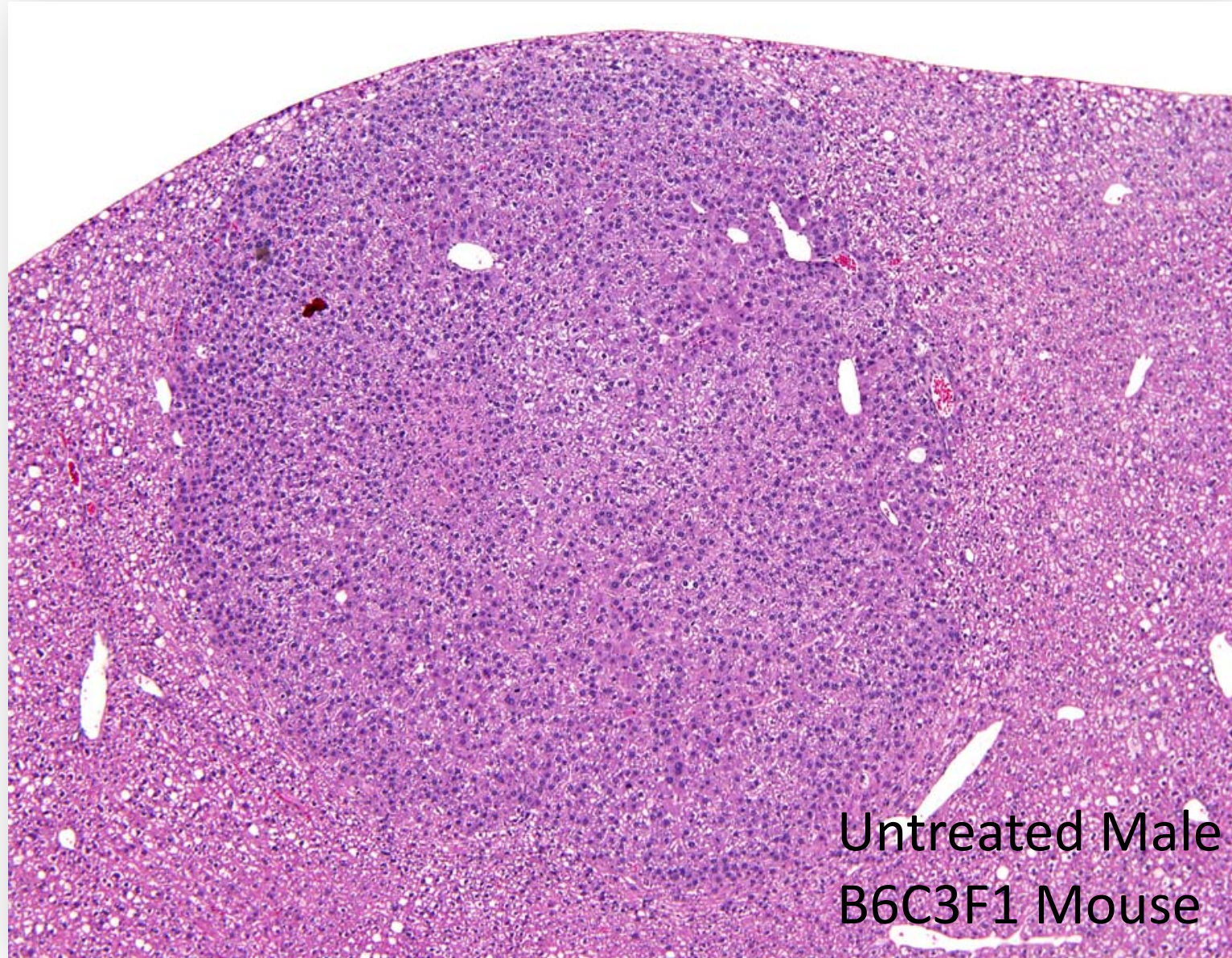
Hepatocellular Adenomas and Carcinomas



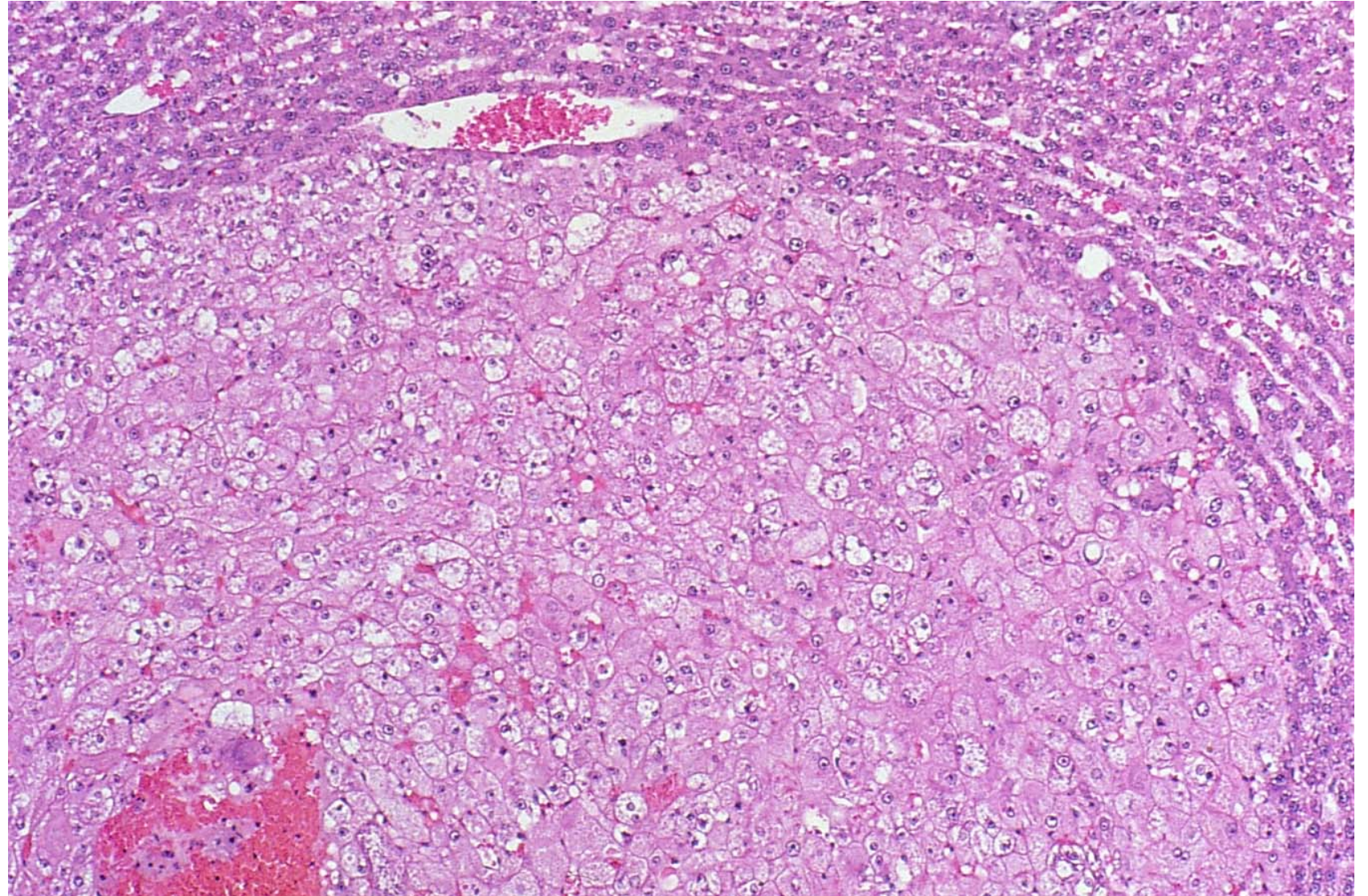
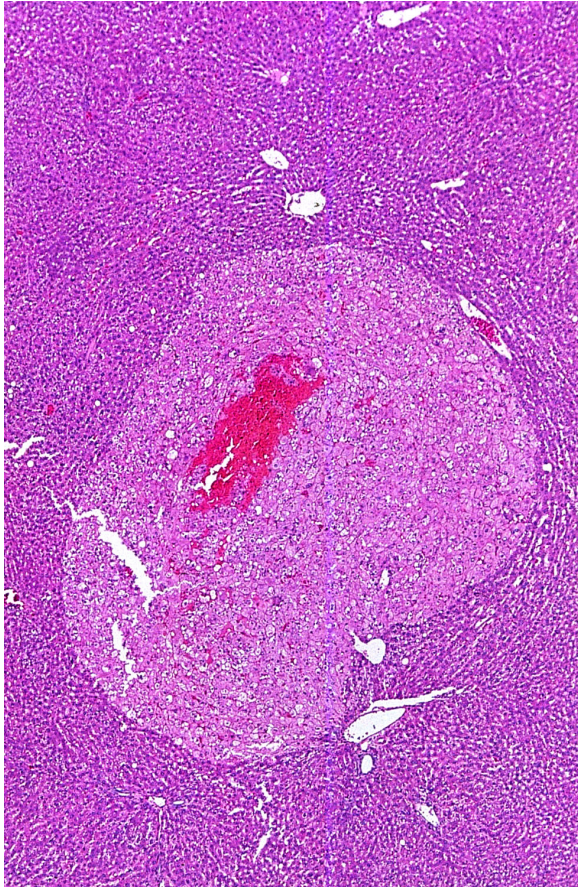
Hepatic Foci of Cellular Alteration



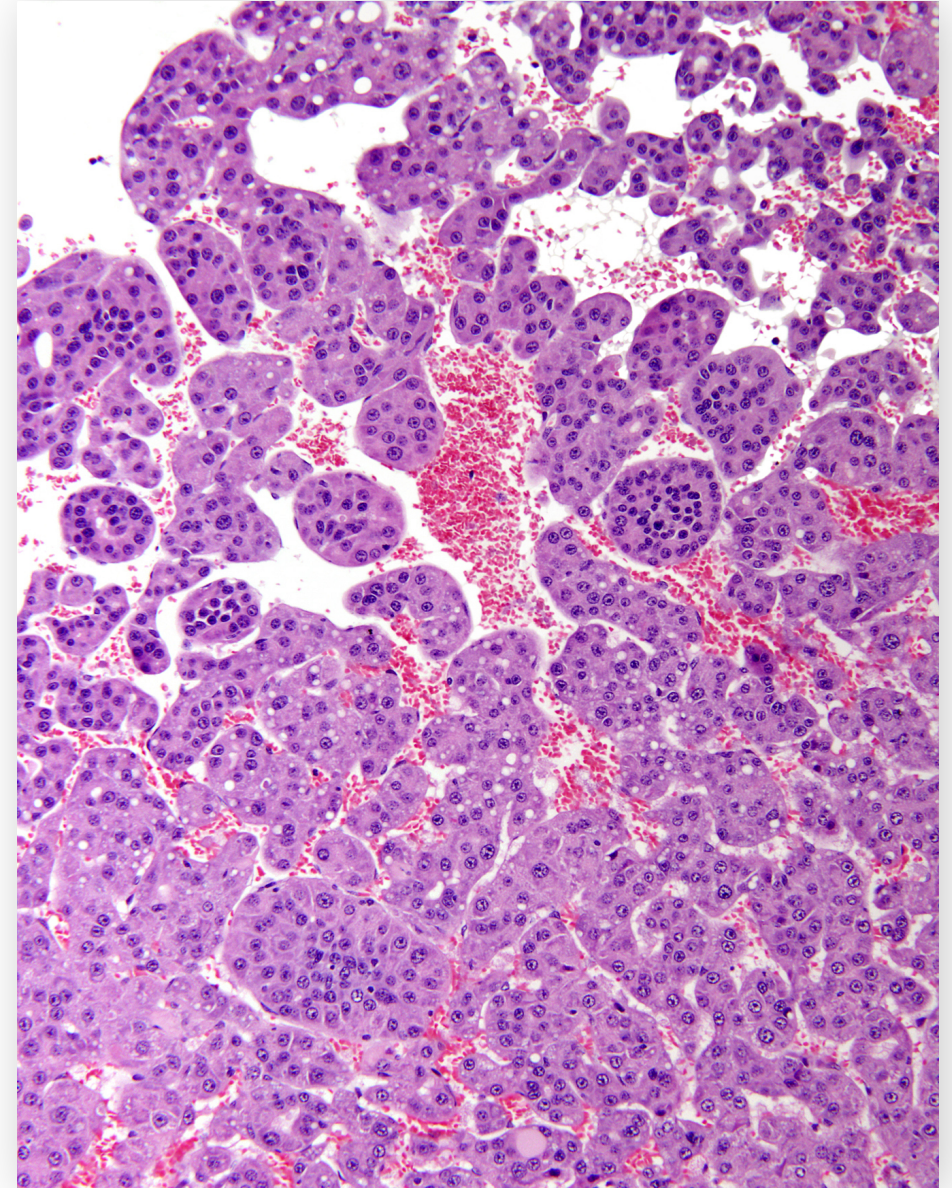
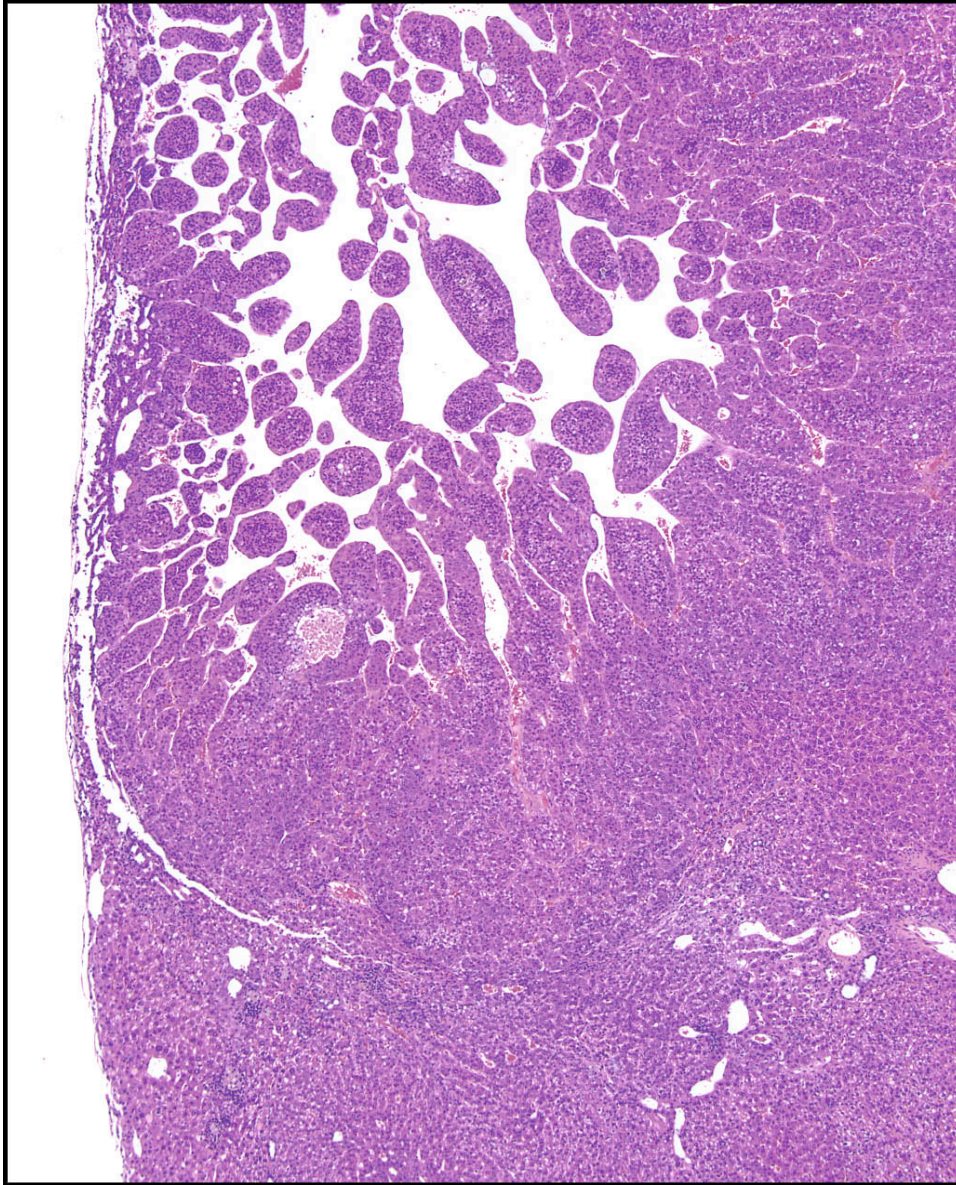
Hepatocellular Adenoma



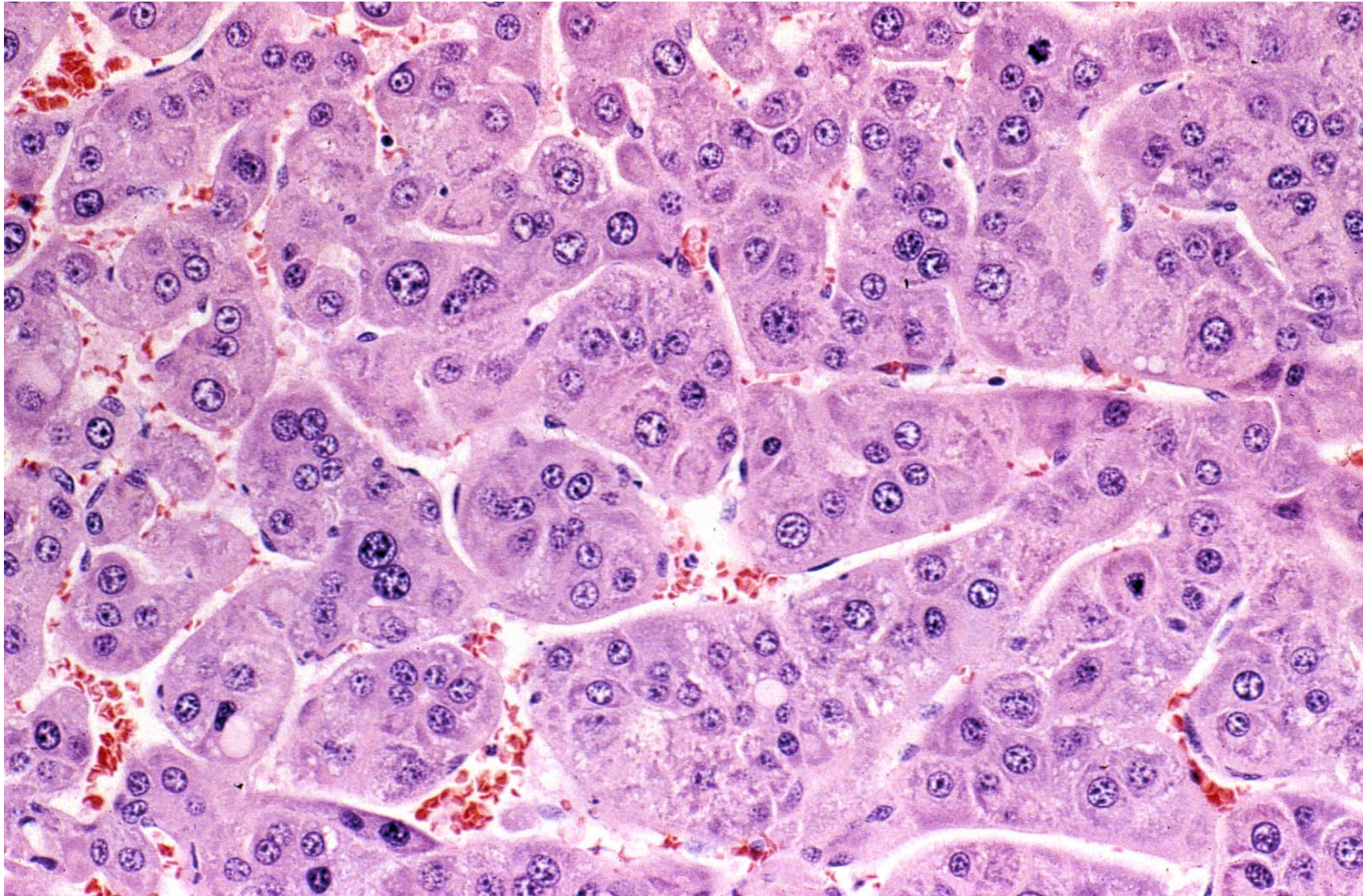
Hepatocellular Adenoma



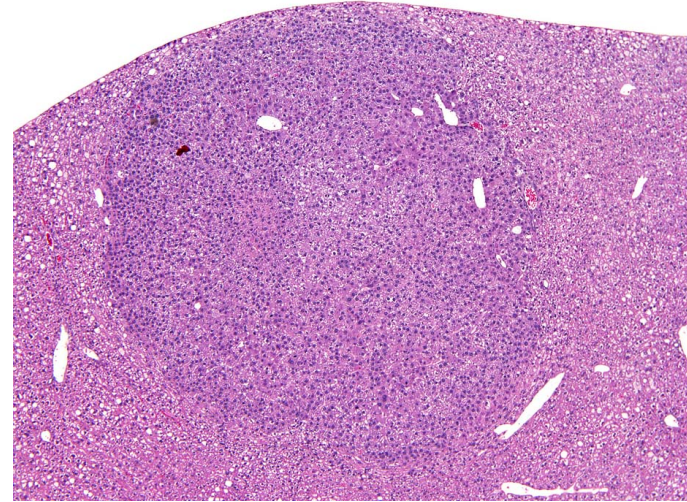
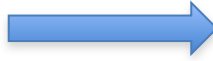
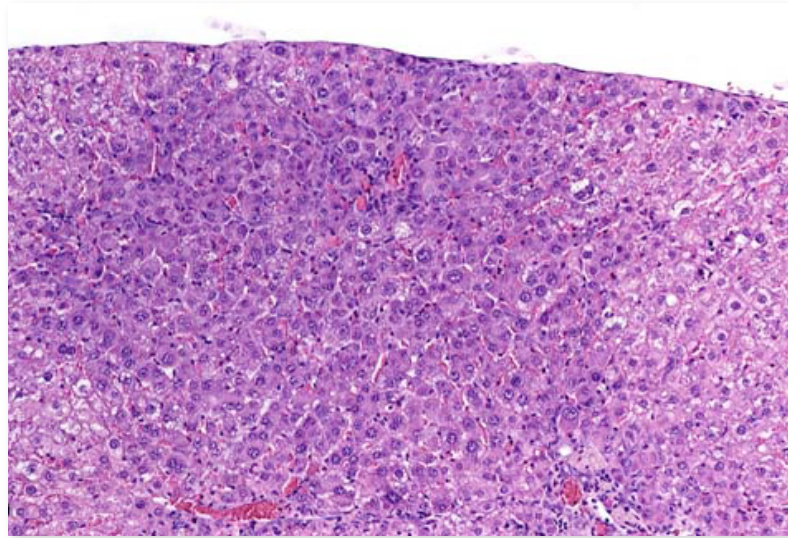
Hepatocellular Carcinoma



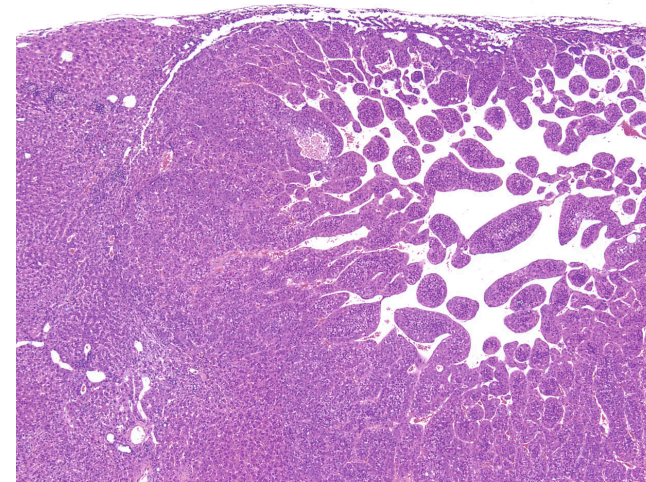
Hepatocellular Carcinoma



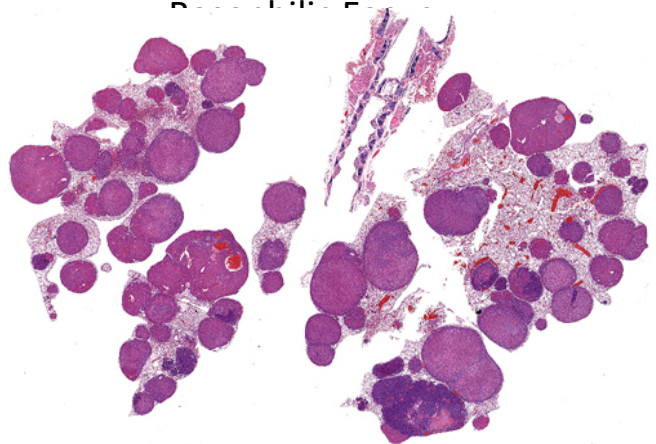
Progression of Proliferative Liver Lesions



Hepatocellular adenoma

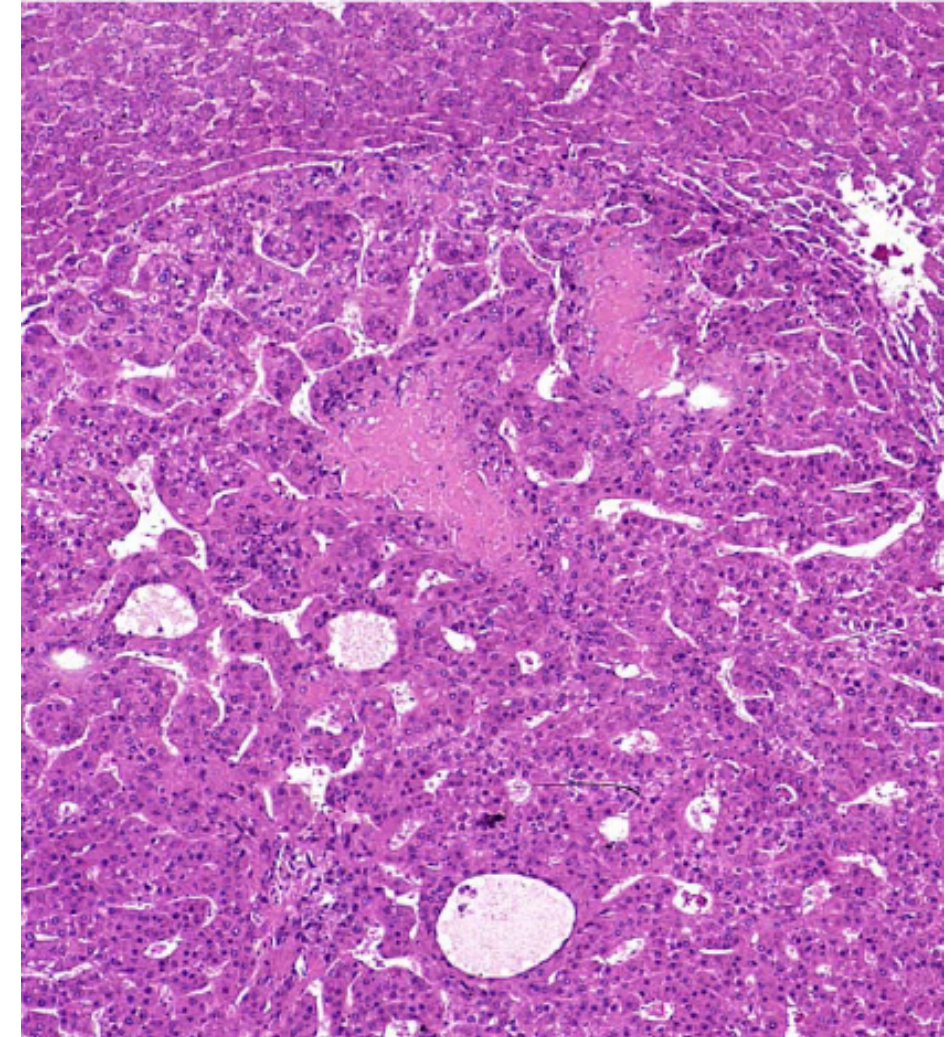
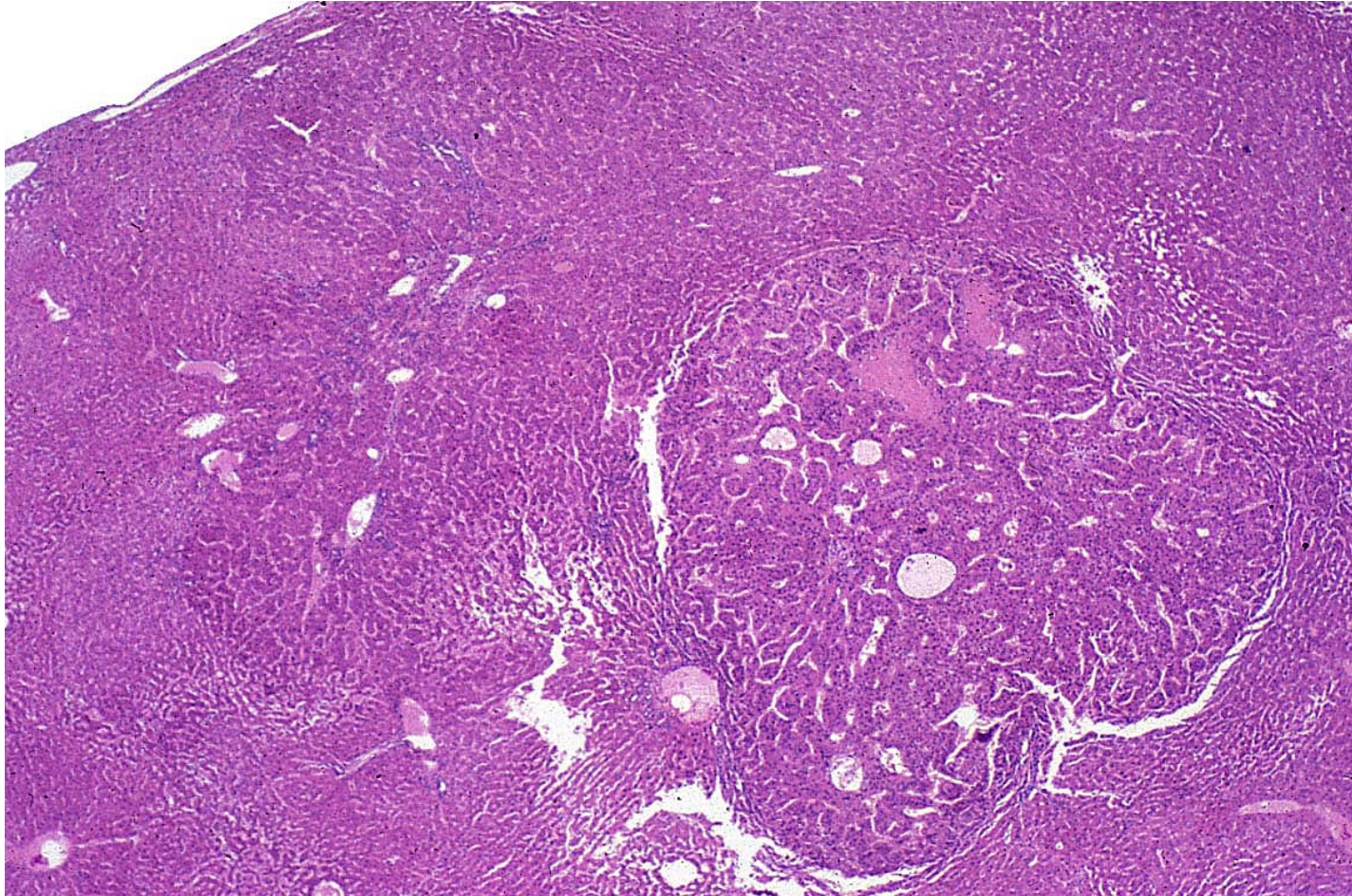


Hepatocellular carcinoma

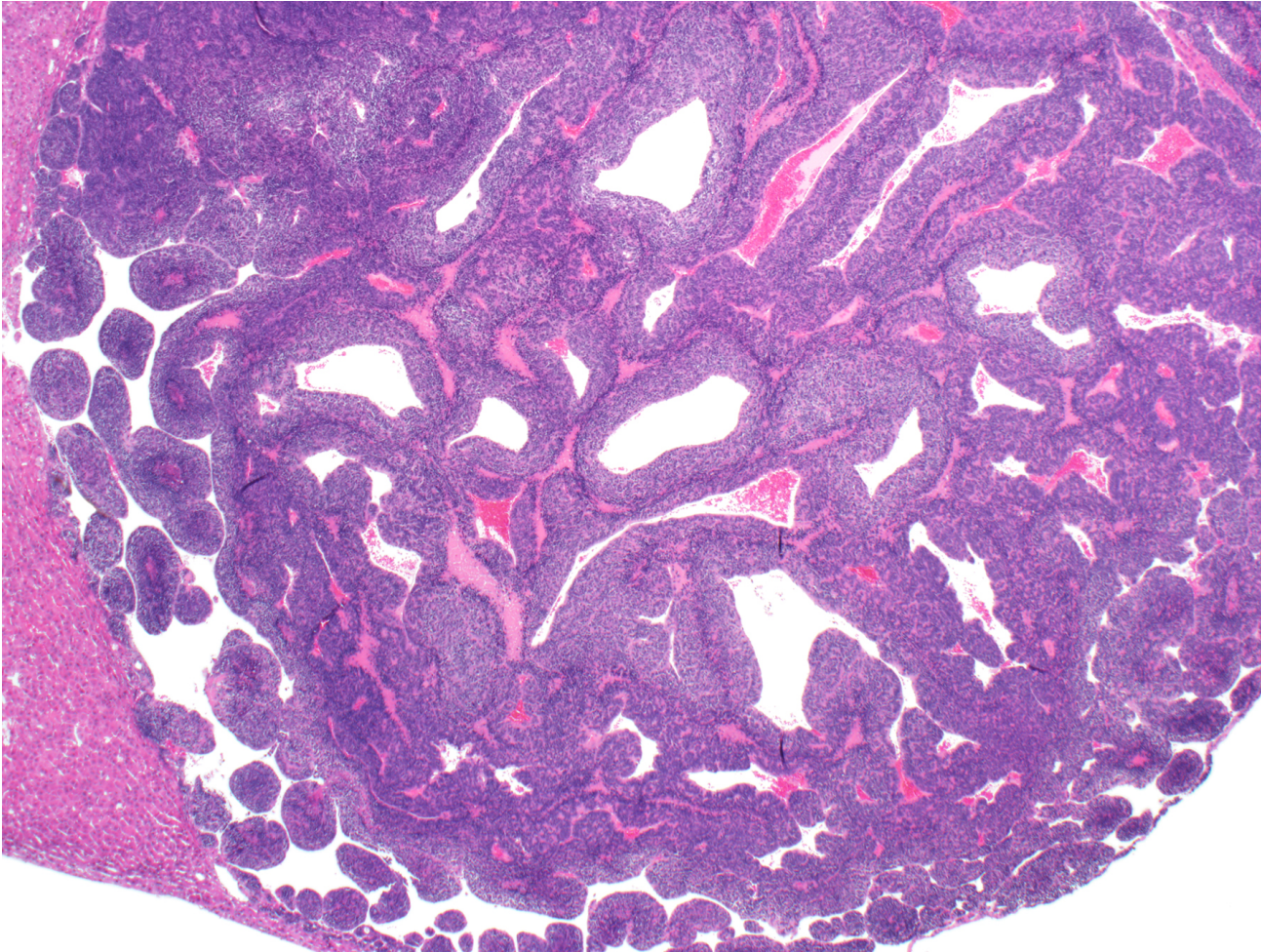


Metastatic carcinoma

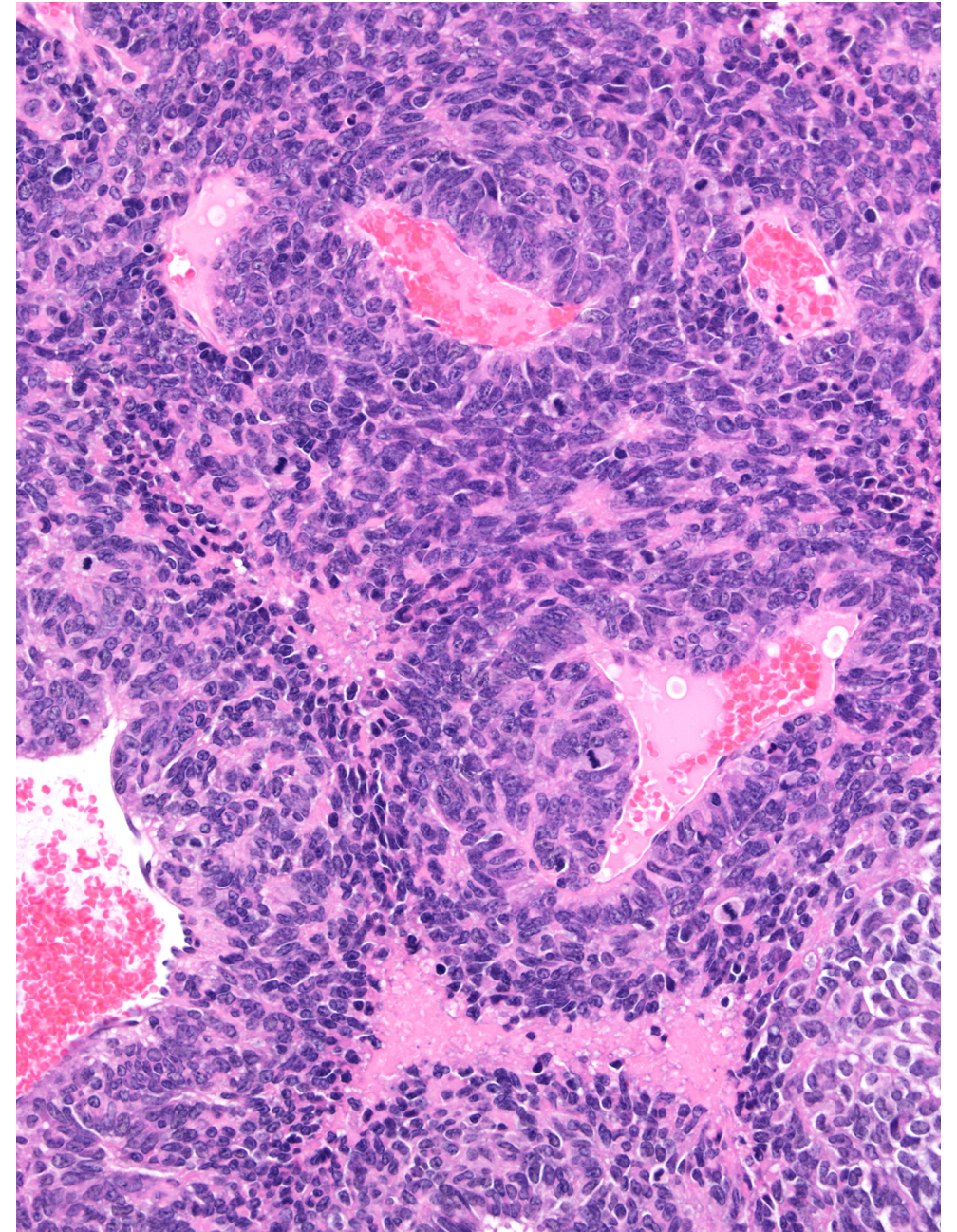
Carcinoma Arising in Adenoma



Hepatoblastoma

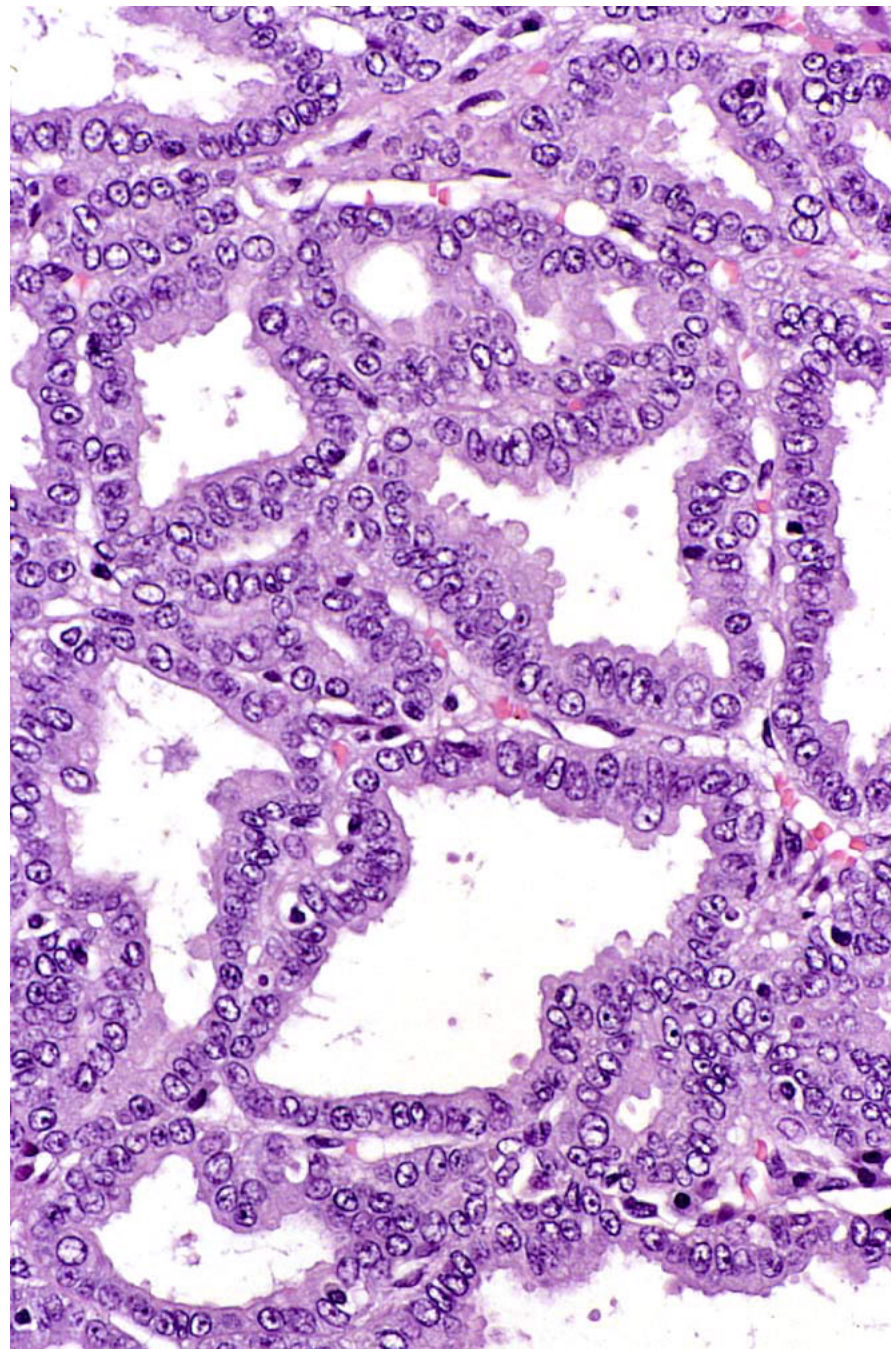
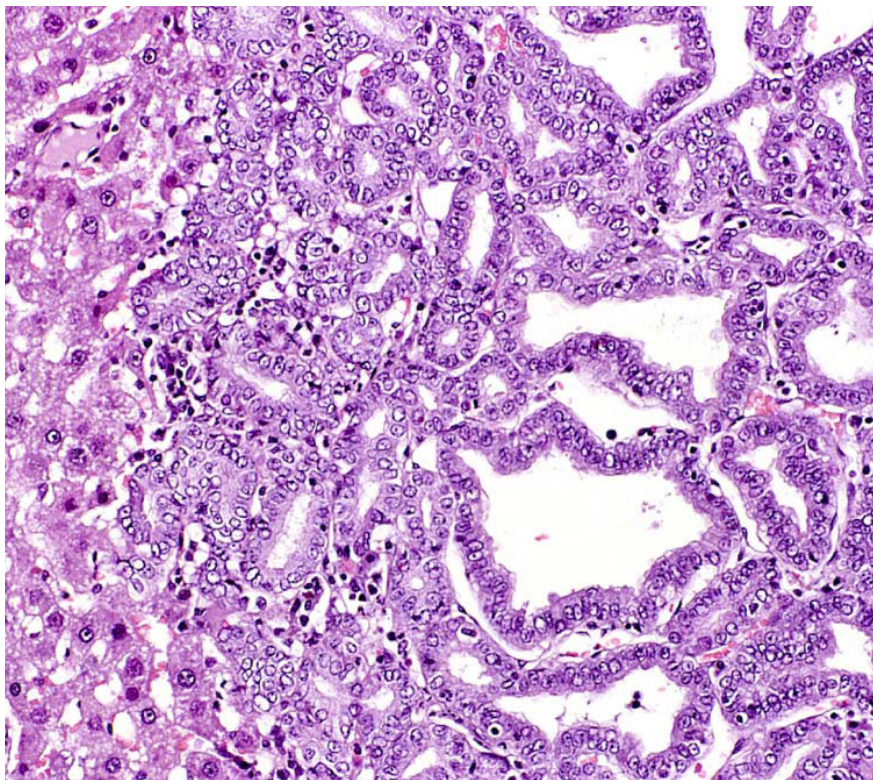
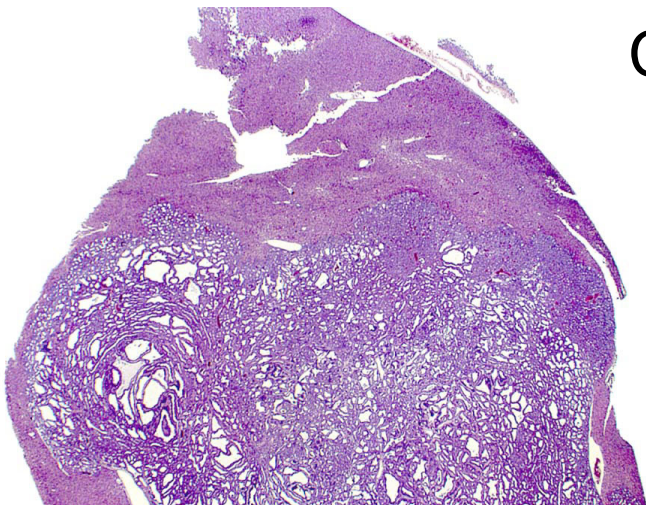


Turusov et al., Tox Path 30(5):580-591 (2002)
(63/140 studies had hepatoblastoma)
(Evaluated 500 hepatoblastomas)

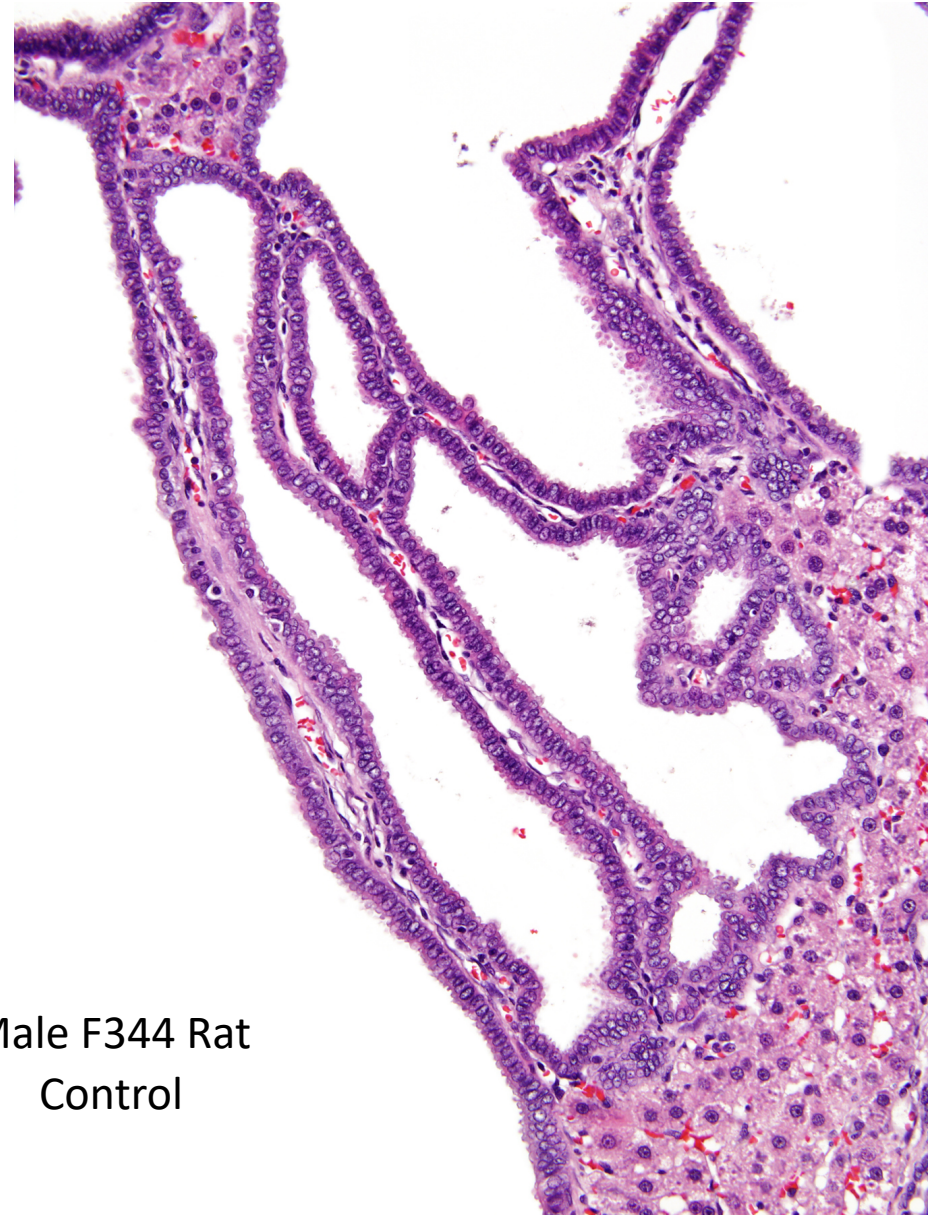
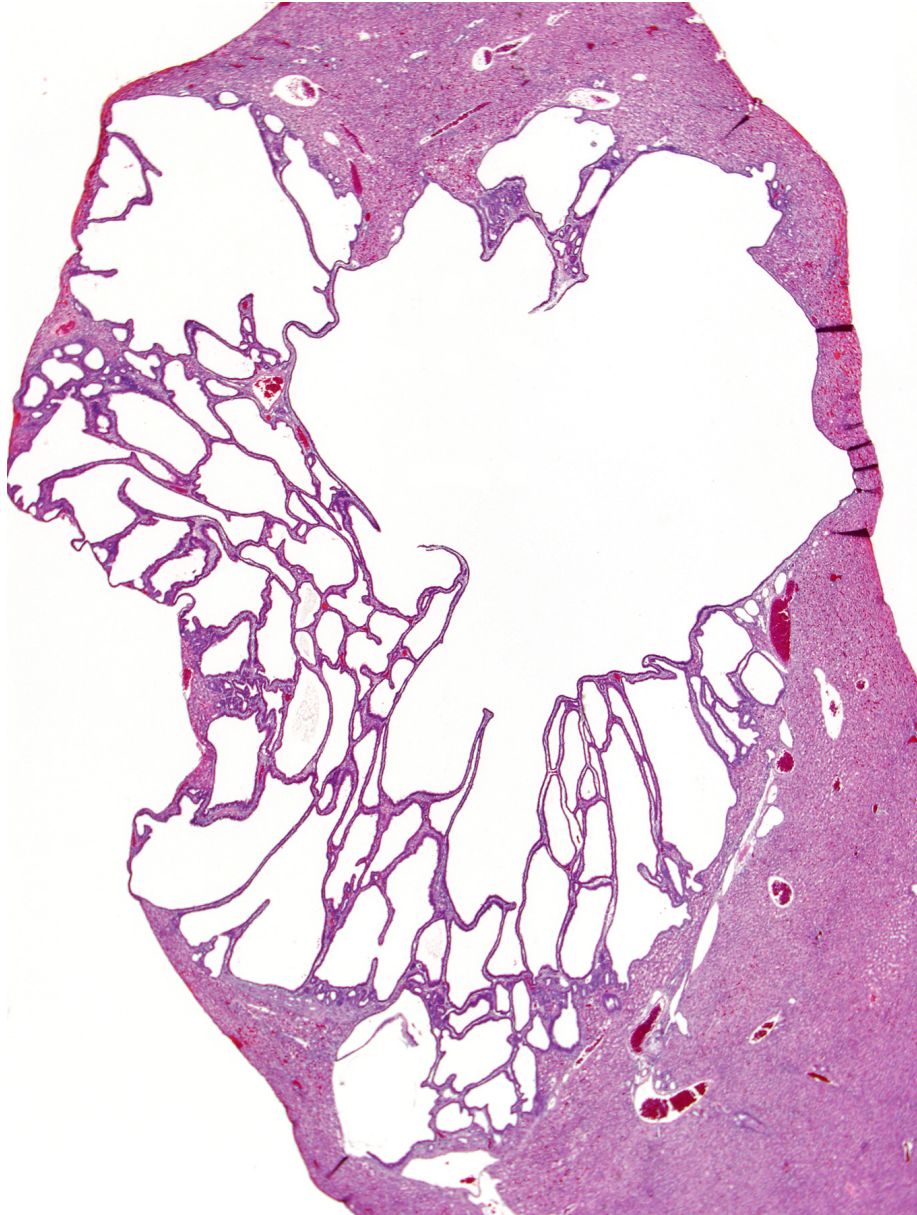


Cholangioma

Sprague Dawley
Male

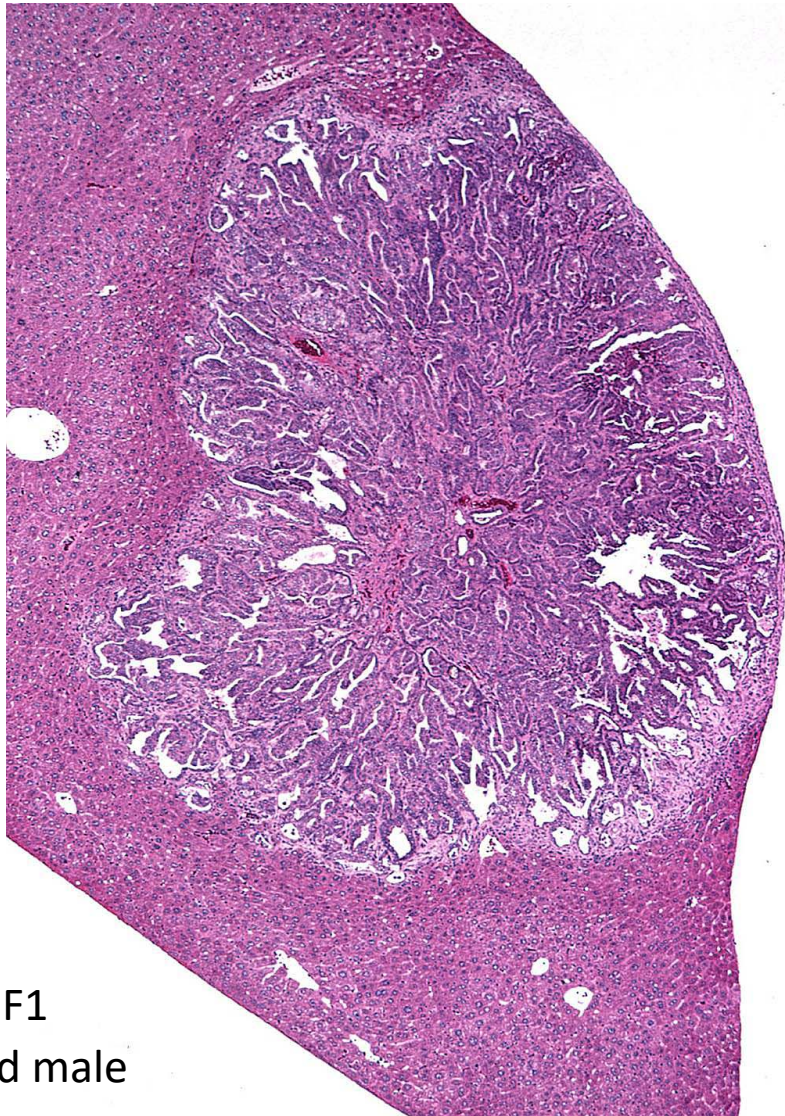


Cystic Cholangioma

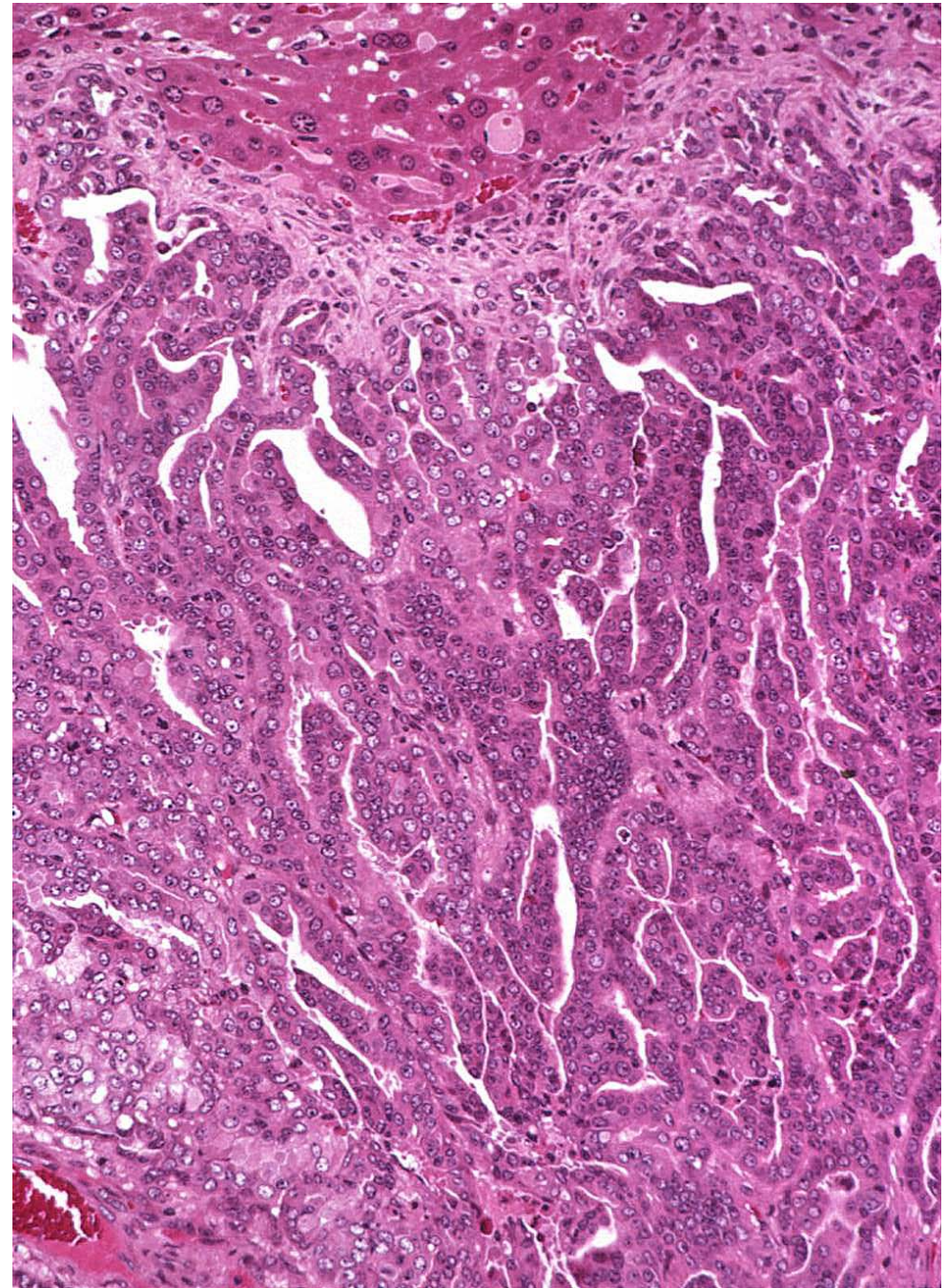


Male F344 Rat
Control

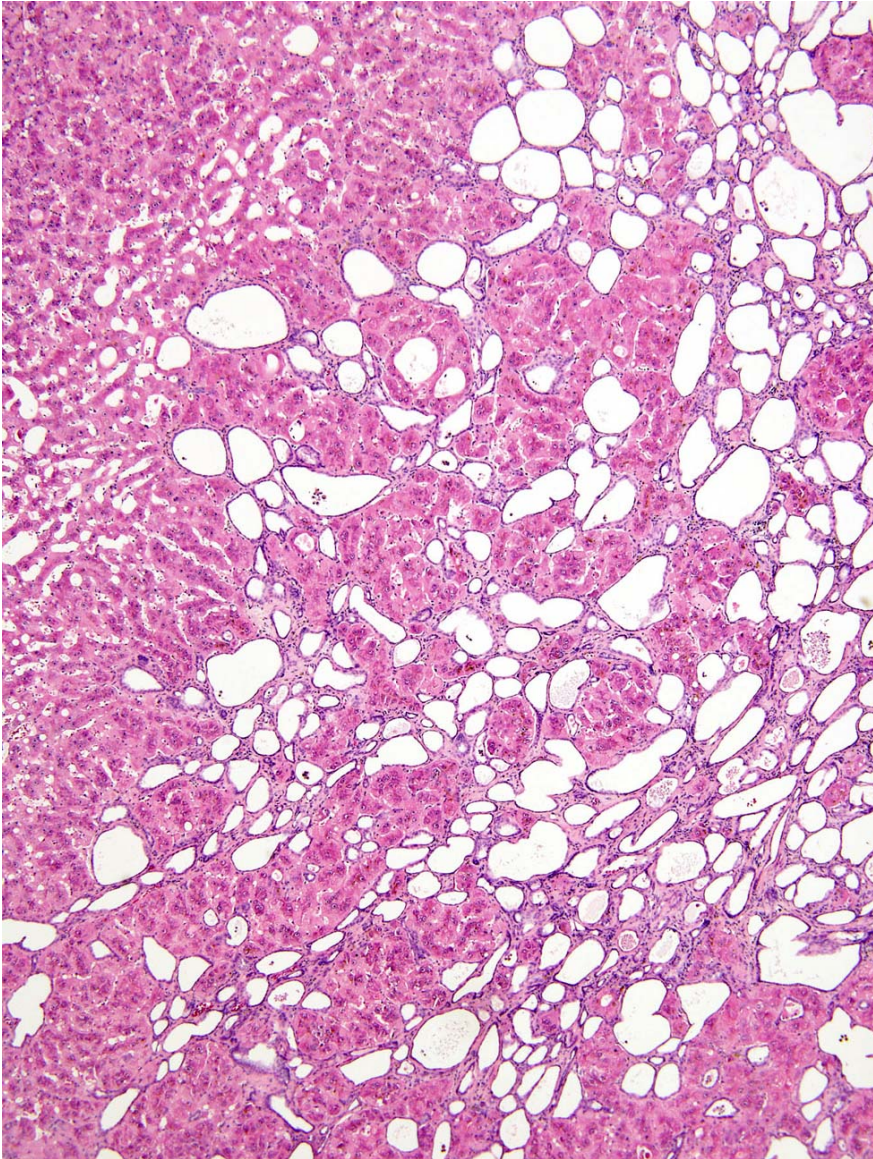
Cholangiocarcinoma



B6C3F1
Untreated male

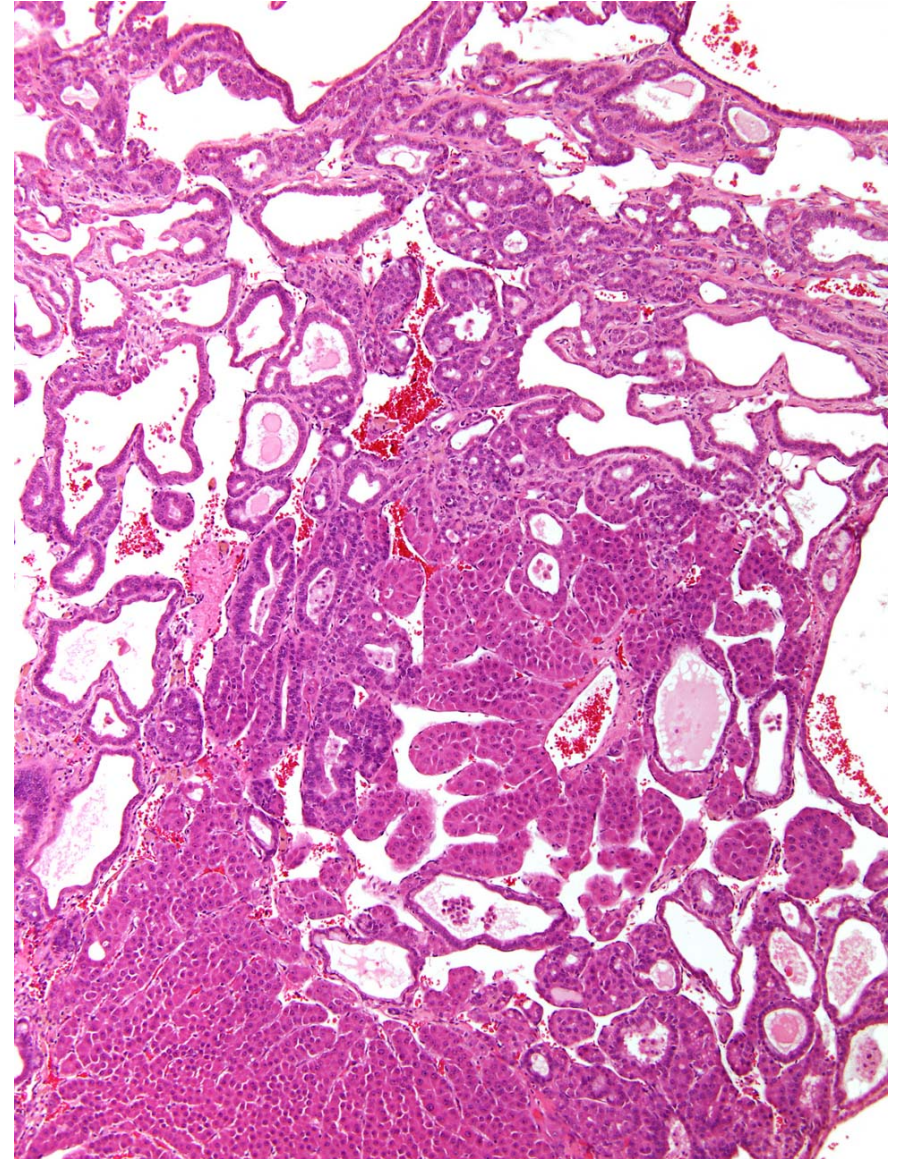


Hepatocholangioma



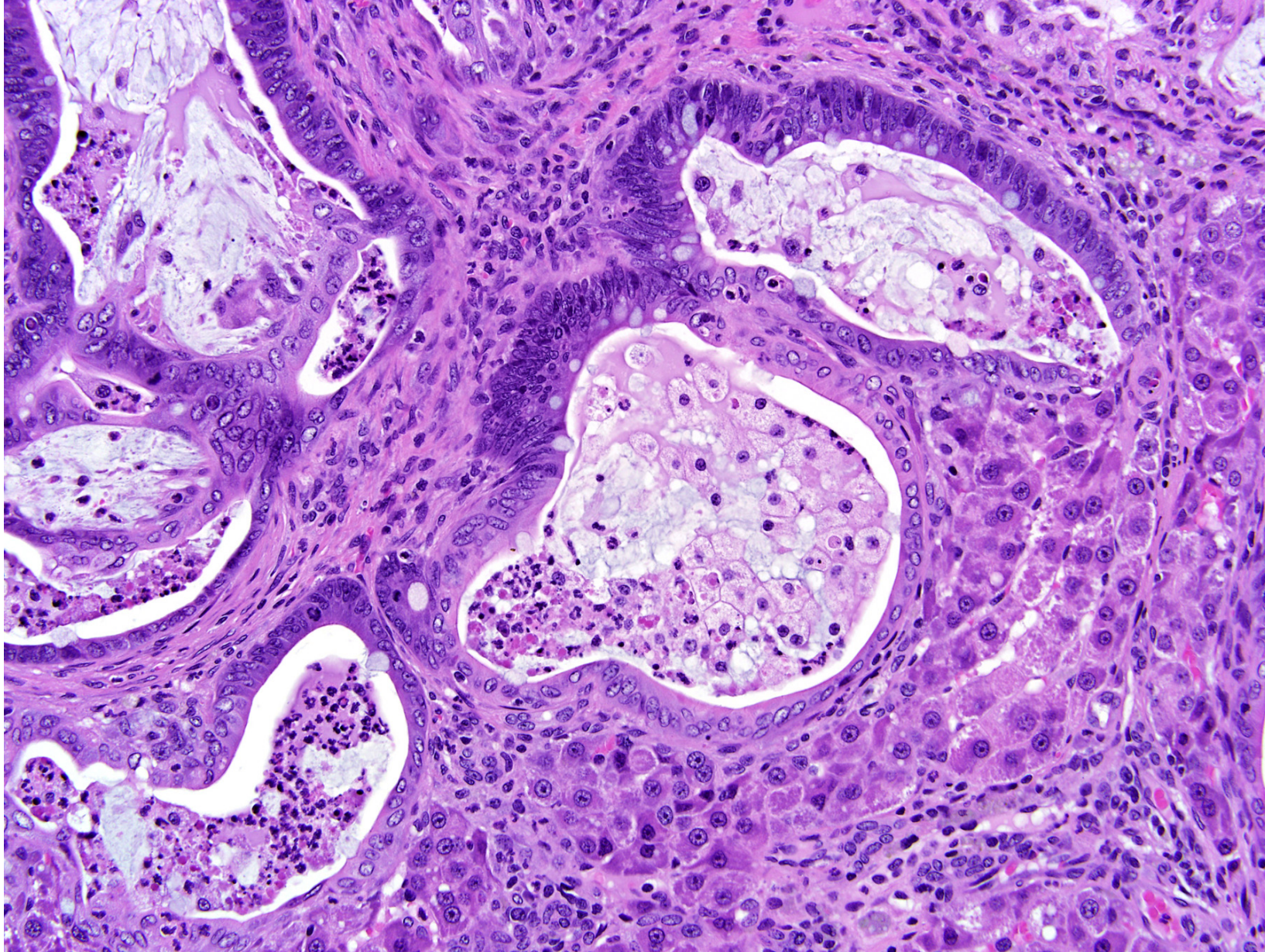
Treated Female Sprague Dawley

Hepatocholangiocarcinoma

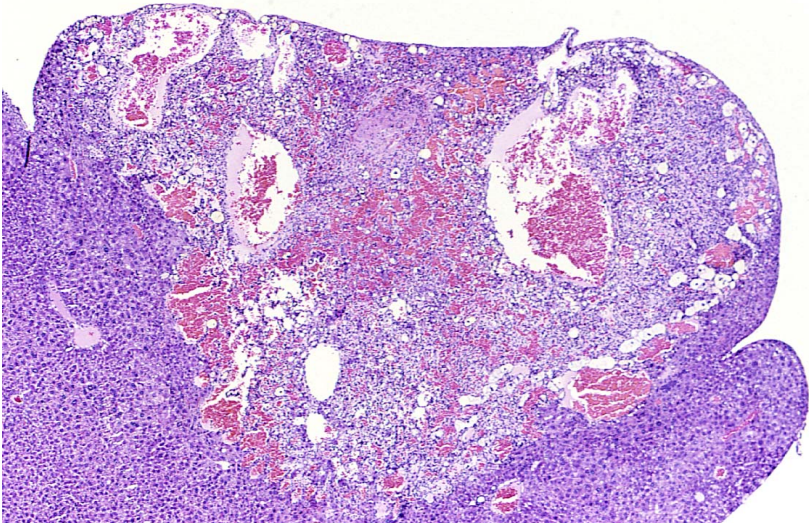


Treated Male F344

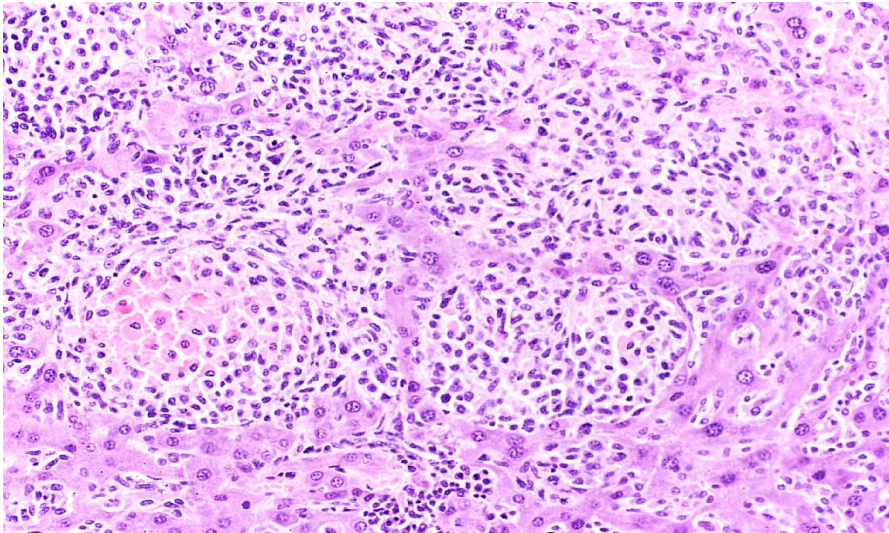
Hepatocholangiocarcinoma with intestinal metaplasia



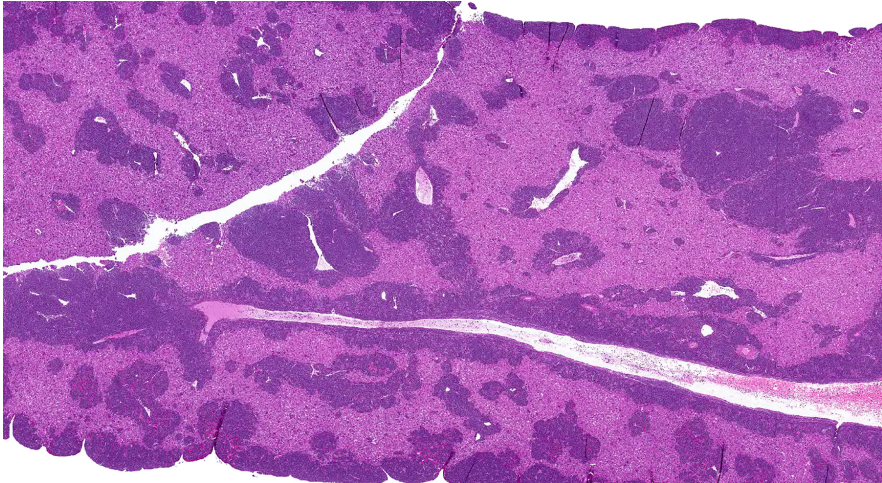
Other types of liver tumors



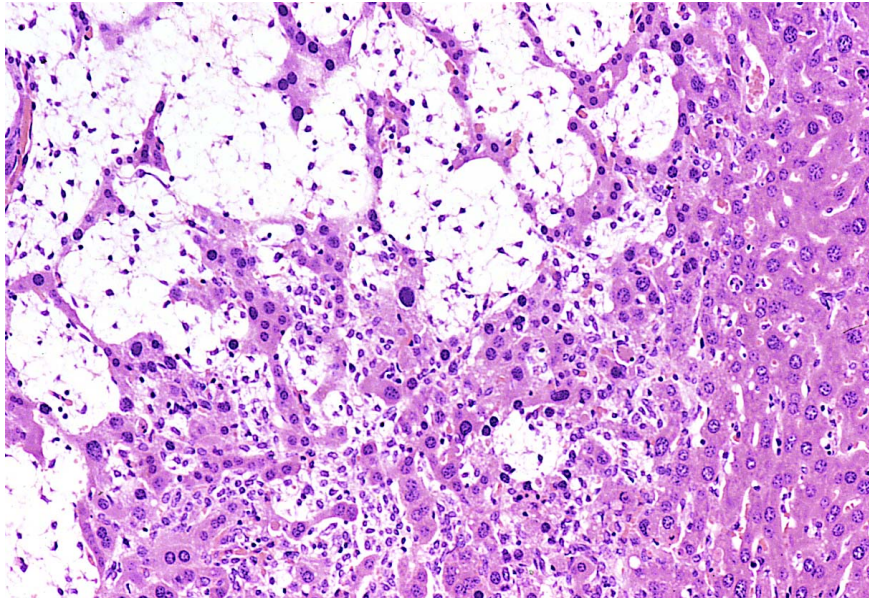
Hemangiosarcoma



Histiocytic sarcoma

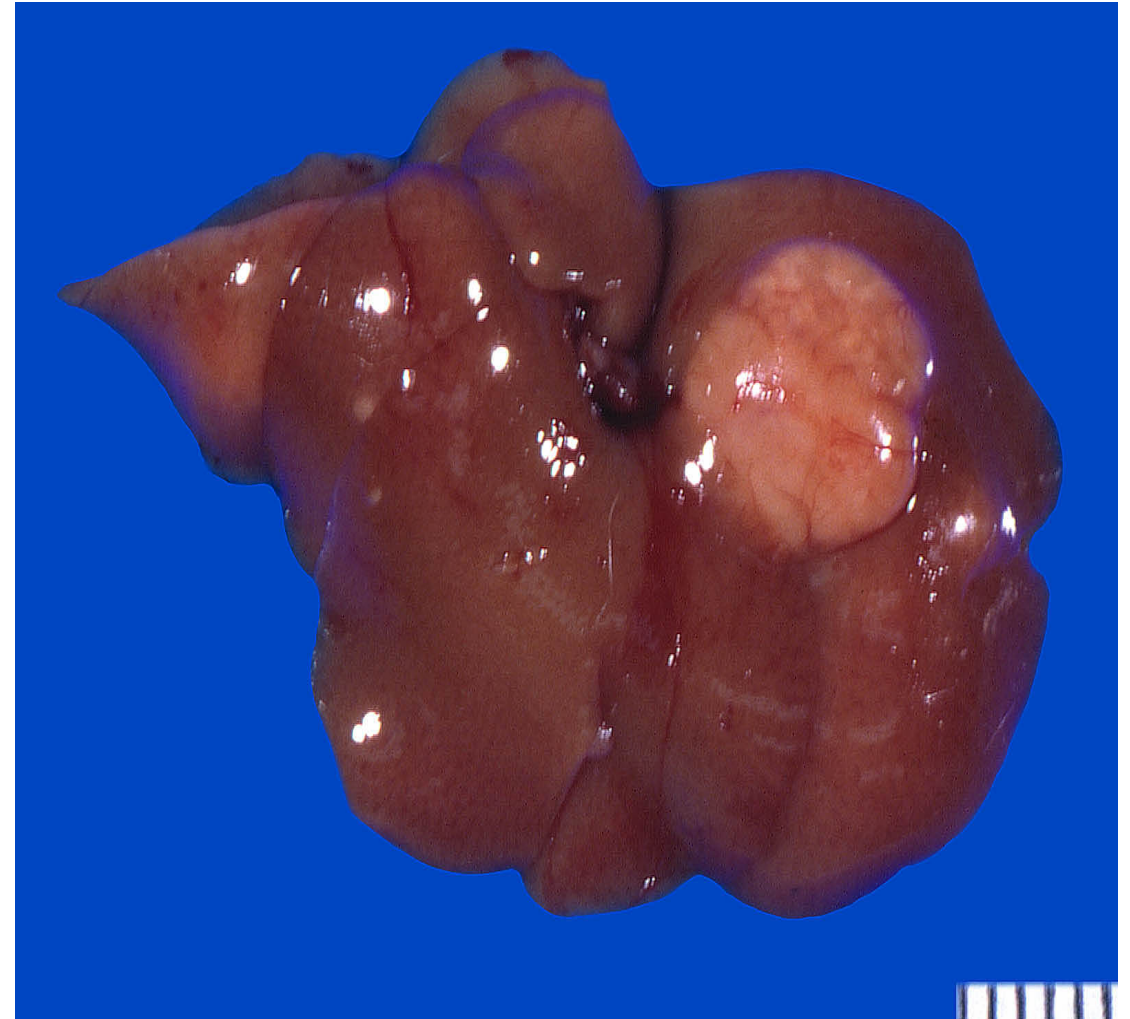


Lymphoma

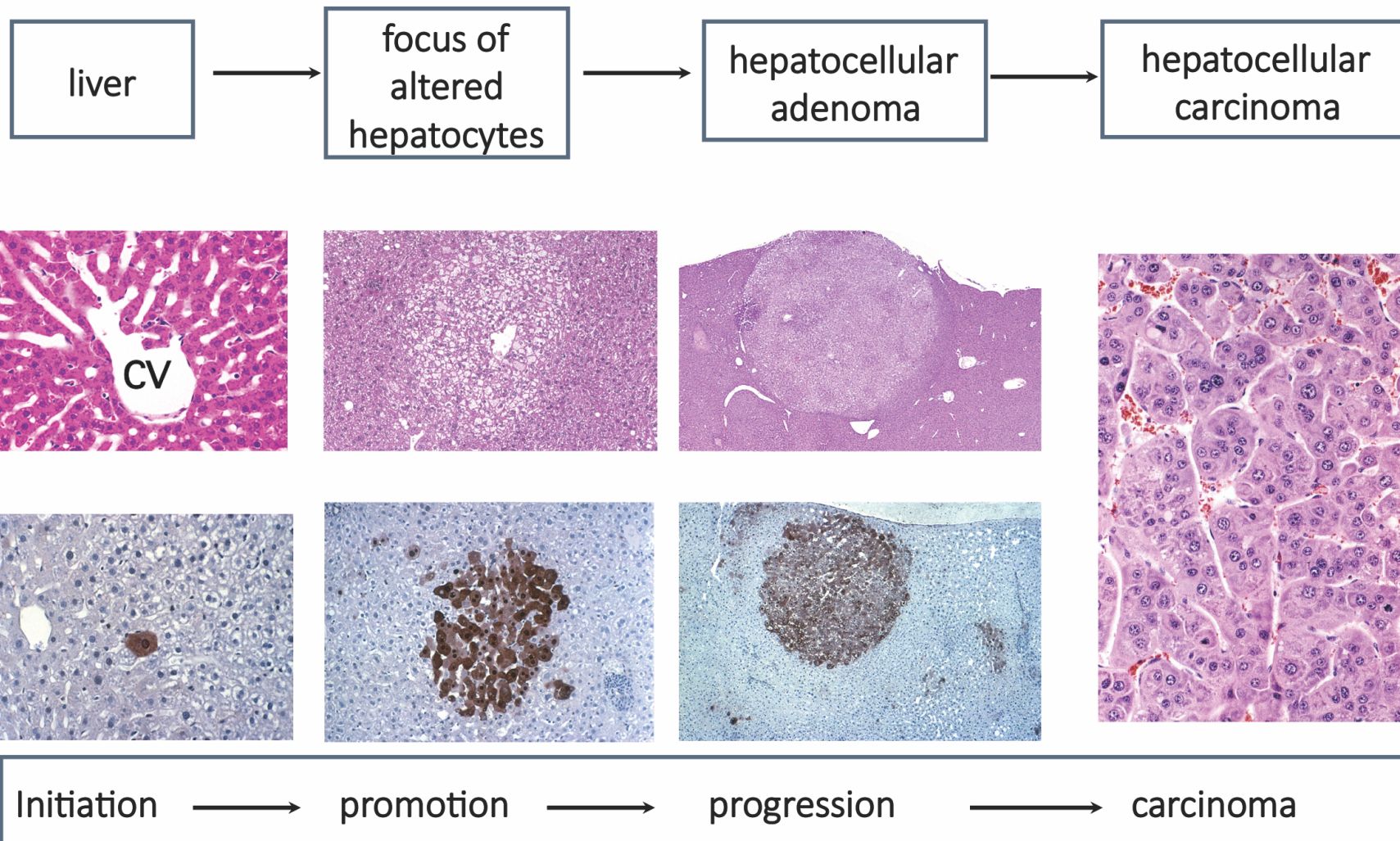


Stellate cell tumor

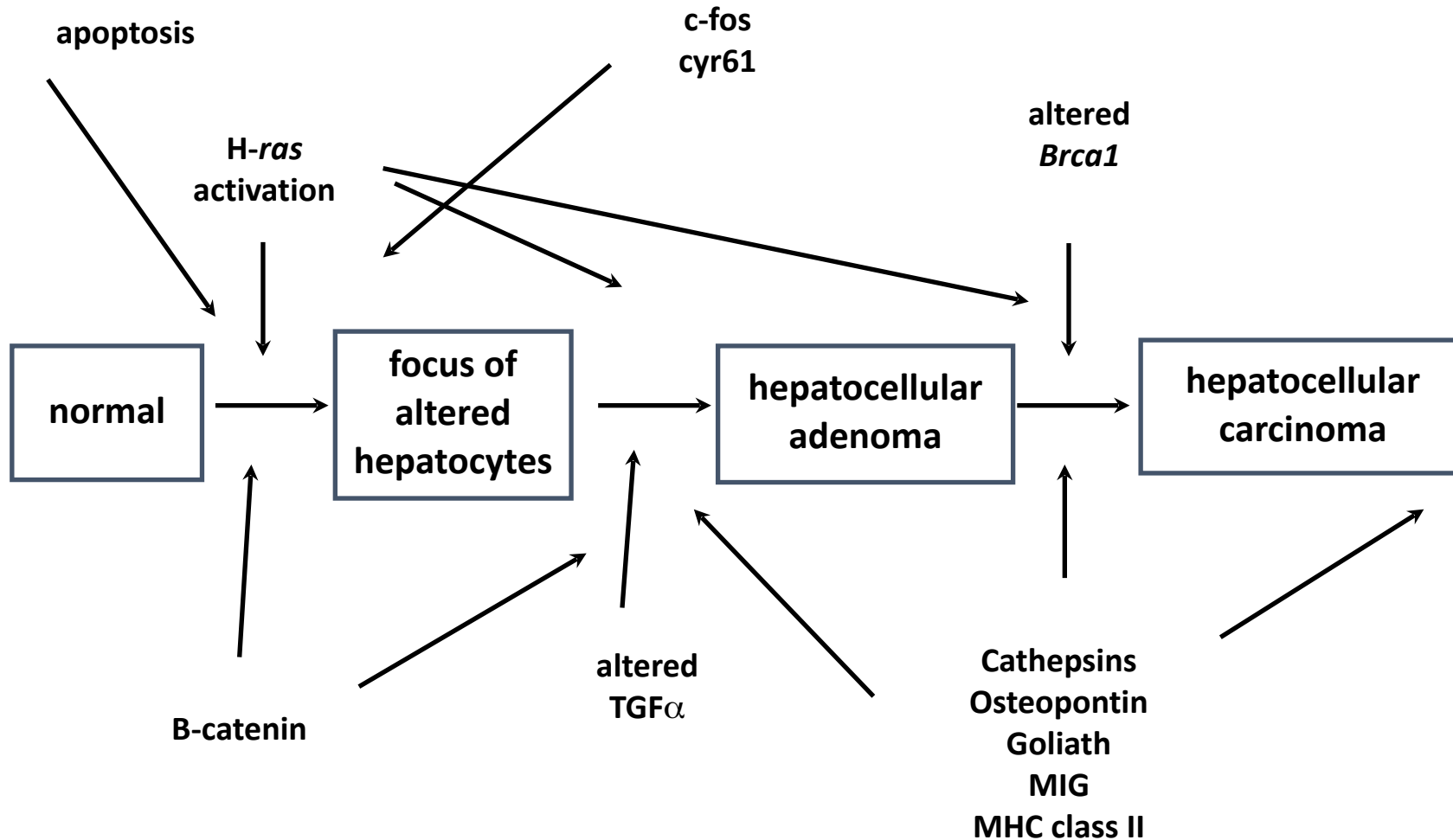
**What have we learned from the conventional bioassay
with respect to liver tumors?**



Multistep hepatocarcinogenesis



Multistage hepatocarcinogenesis



There were and still are some strong opinions
about the significance of rodent bioassays



A Critical Review of the Effectiveness of Rodent Pharmaceutical Carcinogenesis Testing in Predicting for Human Risk

C. L. Alden¹, A. Lynn¹, A. Bourdeau¹, D. Morton², F. D. Sistare³, V. J. Kadambi¹, and L. Silverman¹

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DOI: 10.1177/0192623310363813

Evaluation of Possible Carcinogenic Risk to Humans Based on Liver Tumors in Rodent Assays: The Two-Year Bioassay Is No Longer Necessary

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ABSTRACT

The PPARα-dependent rodent liver tumor response is not relevant to humans: addressing misconceptions

J. Christopher Corton , Jeffrey M. Peters & James E. Klaunig

Archives of Toxicology 92, 83–119 (2018) | Cite this article

Review and Evaluation of the NCI/NTP Carcinogenesis Bioassays*

GIRARD H. HOTTENDORF AND IRWIN J. PACHTER

Bristol-Myers Company, Pharmaceutical Research and Development
Division, Syracuse, NY 13221

The Limits of Two-Year Bioassay Exposure Regimens for Identifying Chemical Carcinogens

James Huff,¹ Michael F. Jacobson,² and Devra Lee Davis³

¹National Institute of Environmental Health Sciences, National Institutes of Health, Department of Health and Human Services, Research Triangle Park, North Carolina, USA; ²Center for Science in the Public Interest, Washington, DC, USA; ³Center for Environmental Oncology, University of Pittsburgh Cancer Institute, Department of Epidemiology, Graduate School of Public Health, Pittsburgh, Pennsylvania, USA

JOURNAL ARTICLE

Goodbye to the bioassay

Jay I. Goodman

Toxicology Research, Volume 7, Issue 4, July 2018, Pages 558–564,

<https://doi.org/10.1039/c8tx00004b>

Published: 13 February 2018 Article history ▾

J Toxicol Pathol 2007; 20: 13–19

Review

The Two-Year Rodent Carcinogenesis Bioassay — Will It Survive?

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¹Comparative Medicine Branch, Natio
Bethesda, Maryland 20892–8135 US

Evaluation of the utility of the lifetime mouse bioassay in the identification of cancer hazards for humans

Thomas G. Osimitz , , Wiebke Droege , , Alan R. Boobis , , Brian G. Lake ,

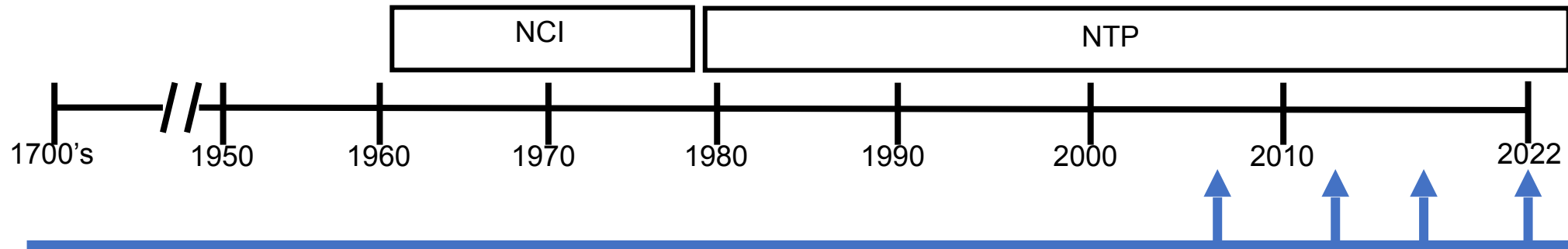
Human relevance of rodent liver tumors: Key insights from a Toxicology Forum workshop on nongenotoxic modes of action

Susan P. Felter , , Jennifer E. Foreman , Alan Boobis , J. Christopher Corton ,
Adriana M. Doi , Lynn Flowers , Jay Goodman , Lynne T. Haber , Abigail Jacobs ,
James E. Klaunig , Angela M. Lynch , Jonathan Moggs , Arun Pandiri

Mode of Action in Relevance of Rodent Liver Tumors to Human Cancer Risk

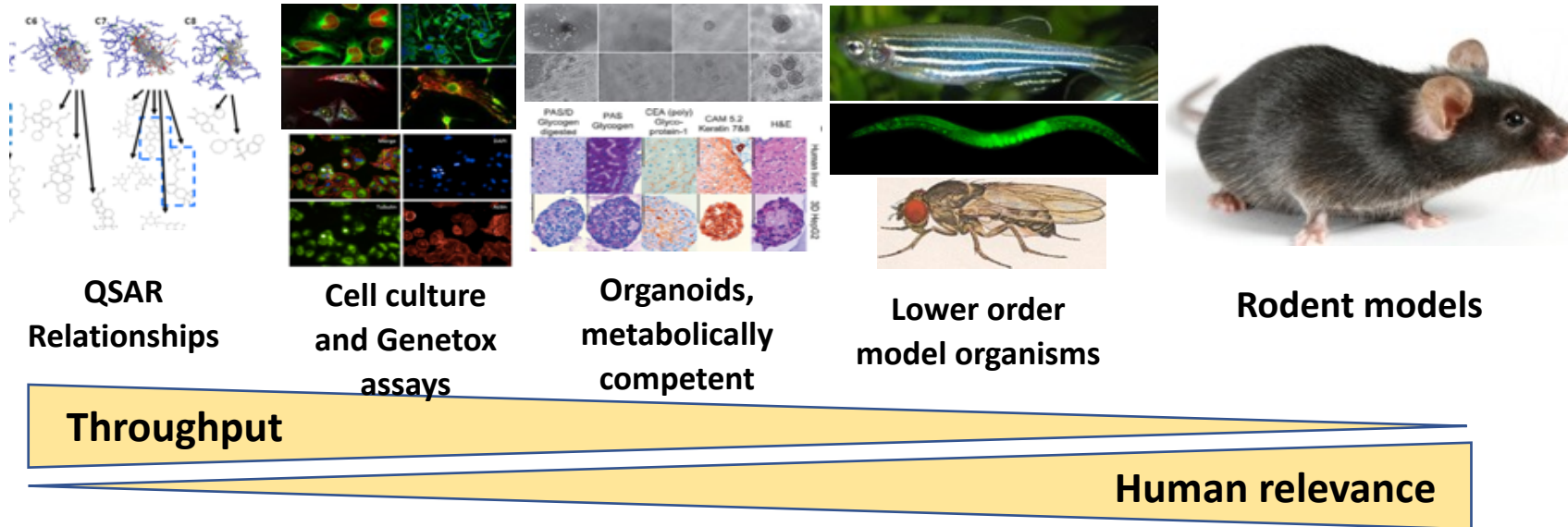
Michael P. Holsapple,^{*,1} Henri C. Pitot,[†] Samuel H. Cohen,[‡] Alan R. Boobis,[§] James E. Klaunig,[¶]
Timothy Pastoor,^{||} Vicki L. Dellarco,^{|||} and Yvonne P. Dragan^{||||}

Mechanisms associated with bioassay tumor responses



- Nuclear receptor activation
 - CAR/PXR, AhR, PPAR-a
- Cytotoxicity and regenerative hyperplasia
- Endocrine modifiers
- Epigenetic modifiers
- Mitogen/tumor promoter
- Inflammation
- Oxidative stress
- Hormonal perturbation
- Immunosuppression
- Suppression of apoptosis

Contemporary NTP efforts



- Core set of mechanistic assays
 - DNA repair & reactivity
 - Receptor-mediated assays
 - Intercellular communication
 - Enzyme induction
 - Cell cycle perturbations
 - Endocrine disruption
 - Effects on methylation
 - Oxidative stress
 - Immunosuppression

- Other contemporary investigative approaches
 - NEGCARC (Genotoxicity, endocrine, histopathology) for pharmaceuticals
 - Tox 21 & high throughput screening assays
 - Genomics, proteomics, metabonomics
 - Mutations in cancer genes
 - Structure activity relationships
 - Epigenetic changes
 - Adverse outcome pathway/MOA

Prechronic liver lesions as predictors of liver carcinogenicity*

B6C3F1 Mouse

- 25 of the 27 (92%) liver tumor positive studies were correctly identified in 90-day studies based on combination of liver hypertrophy, cytomegaly, necrosis and increased liver weight ($p < 0.001$)
- 18 false positives

F344 Rat

- 7 of 11 (64%) liver tumor positive studies were correctly identified in 90-day studies based on combination of liver hypertrophy, cytomegaly, and necrosis ($p < 0.01$)
- 16 false positives

*Based on examination of 83 B6C3F1 and 87 F344 90-day studies with corresponding 2-year studies

